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## Capturing biological activity in natural product fragments by chemical synthesis

Crane, Erika A ; Gademann, Karl

**Abstract:** Natural products have had an immense influence on science and have directly led to the introduction of many drugs. Organic chemistry, and its unique ability to tailor natural products through synthesis, provides an extraordinary approach to unlock the full potential of natural products. In this Review, an approach based on natural product derived fragments is presented that can successfully address some of the current challenges in drug discovery. These fragments often display significantly reduced molecular weights, reduced structural complexity, a reduced number of synthetic steps, while retaining or even improving key biological parameters such as potency or selectivity. Examples from various stages of the drug development process up to the clinic are presented. In addition, this process can be leveraged by recent developments such as genome mining, antibody–drug conjugates, and computational approaches. All these concepts have the potential to identify the next generation of drug candidates inspired by natural products.

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## Drug Discovery

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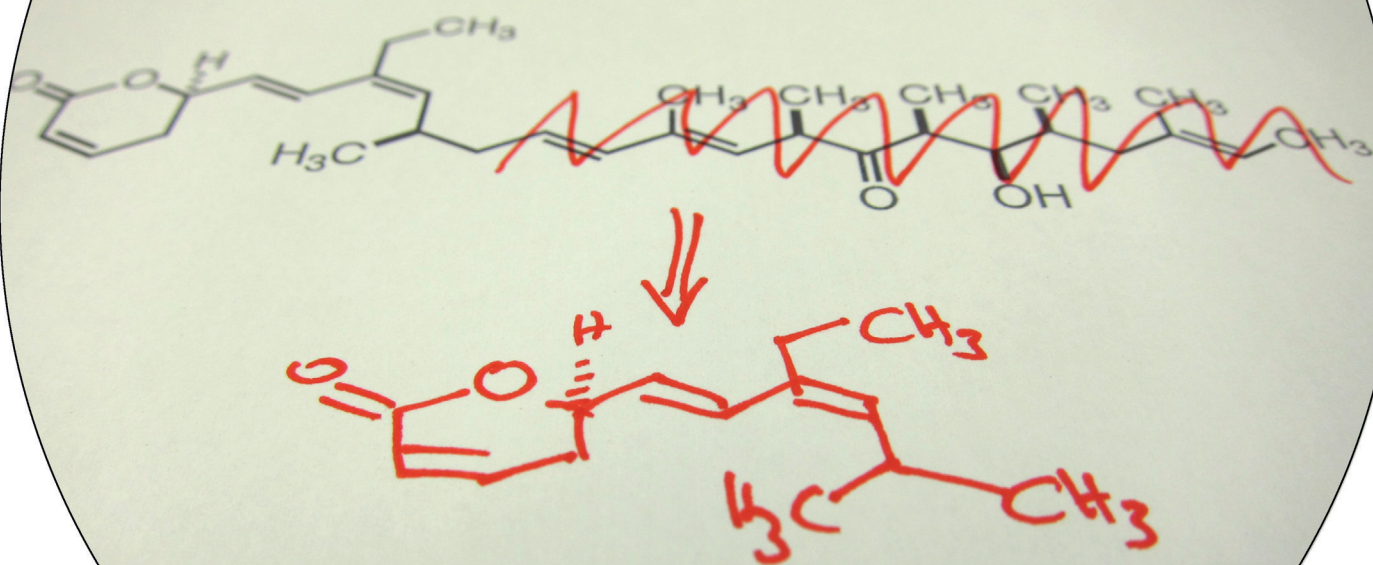
# Capturing Biological Activity in Natural Product Fragments by Chemical Synthesis

*Erika A. Crane and Karl Gademann\**

## Keywords:

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organic synthesis

Reduce  
to the Max



Natural Product Fragments

**N**atural products have had an immense influence on science and have directly led to the introduction of many drugs. Organic chemistry, and its unique ability to tailor natural products through synthesis, provides an extraordinary approach to unlock the full potential of natural products. In this Review, an approach based on natural product derived fragments is presented that can successfully address some of the current challenges in drug discovery. These fragments often display significantly reduced molecular weights, reduced structural complexity, a reduced number of synthetic steps, while retaining or even improving key biological parameters such as potency or selectivity. Examples from various stages of the drug development process up to the clinic are presented. In addition, this process can be leveraged by recent developments such as genome mining, antibody–drug conjugates, and computational approaches. All these concepts have the potential to identify the next generation of drug candidates inspired by natural products.

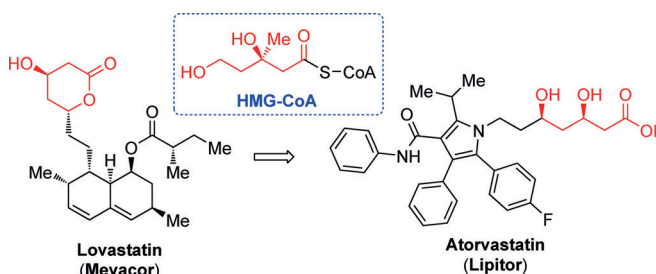
## 1. Introduction

At the advent of research into natural products, organic chemistry was a crucial tool to establish the chemical structure of compounds. As a result, high-yielding, versatile, and specific synthetic methods were developed, which led, with the help of innovative strategies, to the production of natural products in quantities that otherwise would be inaccessible from natural sources and finally enabled more thorough biological evaluation. Once organic chemists could harness the power of synthesis, they were able to modify natural products with an aim to improve the selectivity, potency, stability, and pharmacokinetics. Structure–activity relationship (SAR) properties were investigated by semisynthesis or fragment exchange of natural products, which led to crucial discoveries.

A prime example in this regard has been the development of statins for the treatment of cardiovascular disease, still the leading cause of death worldwide. Although the first generation compounds introduced into the clinic were either identical or closely related to a natural product (lovastatin), the recognition of the crucial 3,5-dihydroxypentanoate scaffold and optimization by medicinal chemistry led to much more improved derivatives, which had a dramatic impact on medicine and society (Figure 1).<sup>[1]</sup> More stringent criteria of “drug likeness” following Lipinski’s “rule of five” and related ADME/pharmacokinetic criteria (ADME = adsorption, distribution, metabolism, excretion), along with the introduction of high-throughput screening (HTS), led to a phasing out of natural products in the lead generation process in drug discovery in the late 1990s. However, in the past 25 years, this paradigm pendent on combinatorial chemistry for medicinal chemistry research has barely provided sufficient numbers of innovative and effective new drugs in Pharma R&D. Interestingly, half of all the new chemical entities (NCEs) introduced over the last 30 years (540 out of 1073) were

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**Figure 1.** Development of HMG-CoA reductase inhibitors for the treatment of cardiovascular disease. CoA = coenzyme A, HMG = 3-hydroxy-3-methylglutaryl.

based on natural products, which underlines their importance for lead generation.<sup>[2]</sup>

Over the last decade, natural products have been experiencing a renaissance in drug discovery; this is based on a number of reasons, in addition to the statistics regarding NCEs presented above. Many studies have revealed striking differences between natural and synthetic compounds, with

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the former being less hydrophobic, having more stereogenic centers, a larger fraction of  $sp^3$ -hybridized carbon atoms, more O and less N, S, and halogen atoms, fewer rotatable bonds, more fused, bridge, or spiro rings, and more solvated hydrogen bond donors and acceptors.<sup>[3]</sup> After all, these compounds produced by nature are a result of millions of years of evolutionary selection and are, hence, biologically prevalidated. All these factors have led to the validation of natural products as prime starting points for drug discovery.

One of the main factors often negatively associated with natural products as lead structures constitutes their limited chemical tractability. The above-mentioned complex architecture, which sometimes requires lengthy synthetic approaches, often disfavors selection of natural products for further study in a research environment that is heavily under pressure in terms of time and financial constraints. It is the goal of this Review to showcase with several examples from early development to the clinic that the structural complexity of natural products can be reduced by the chemical synthesis of smaller fragments (“less is more”, “reduce to the maximum”), while retaining (or sometimes improving) desired biological parameters such as potency or selectivity. This Review is not aimed at being comprehensive,<sup>[4]</sup> but instead should guide the reader through successful design principles and compounds that are at various stages of the development process, from lead structure identification to the clinic. The ultimate goal is shared or complemented by approaches such as “diverted total synthesis”, “chemical or molecular editing”,<sup>[5]</sup> and “function-oriented synthesis” (FOS).<sup>[4a,6]</sup>

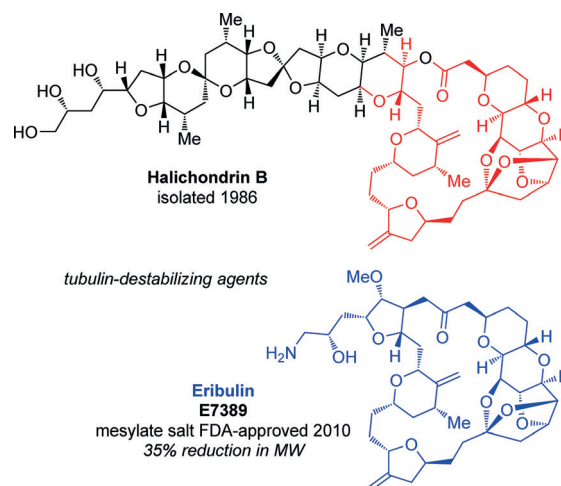
These approaches all present a unique opportunity of retaining desired effects in minimized, biologically active scaffolds as a consequence of “fragment likeness” with a high degree of three dimensional similarity, yet overcome the limitations of parent natural product structures, such as lack of accessibility or viable synthetic approachability. In addition, these natural product derived fragments could add to the discovery process in the context of fragment-based drug discovery.<sup>[7]</sup>

## 2. Natural Product Derived Fragments in Drug Discovery

### 2.1. Anticancer Activity

#### 2.1.1. From Halichondrin B to Eribulin

The polyether macrolide halichondrin B<sup>[8]</sup> was reported in 1986 by Hirata and Uemura (Figure 2).<sup>[9]</sup> It was isolated from the marine sponge *Halichondria okadai* Kadota, which had



**Figure 2.** Structural comparison between halichondrin B and eribulin/ E7389. FDA = U.S. Food and Drug Administration.

been collected off the coast of Japan in the Western Pacific. Later in 1991, Pettit et al. also isolated halichondrin B from an *Axinella* sp. sponge, collected in Palau.<sup>[10]</sup> This marine macrolide showed potent activity against B16 melanoma cells ( $IC_{50} = 0.093 \text{ ng mL}^{-1}$ ) and powerful in vivo inhibition of tumor growth (T/C)<sup>[11]</sup> in mice (in up to 375% of the test group) over the control group and increased the mean survival times for groups with murine B16 melanoma, P388 leukemia, and L1210 leukemia.<sup>[9b]</sup> Later studies revealed that halichondrin B acts as a tubulin-destabilizing agent with a slightly different mechanism of action from other antimitotics, such as the vinca alkaloids.<sup>[8a,12]</sup> More specifically, it inhibits tubulin polymerization and microtubule assembly, tubulin-dependent GTP hydrolysis, nucleotide exchange on



Karl Gademann (born 1972) was educated at the ETH Zürich and Harvard University (PhD with Prof. Dr. Dieter Seebach, post-doctoral studies with Prof. Dr. Eric N. Jacobsen, and habilitation with Prof. Dr. Erick M. Carreira). He started his independent research at the EPFL Lausanne, before moving to the University of Basel in 2010 as a full professor. He has received several awards, including the Novartis Early Career Award, the National Latsis Prize, the Ruzicka Medal, and the European Young Investigator Award. In August 2015 he moved to the University of Zürich.



Erika Crane was born in Kettering, Ohio (USA) in 1985. Her research career began as an undergraduate with Prof. Robert S. Coleman at The Ohio State University. She received her BS in Biochemistry from Ohio Northern University in 2007, and then her PhD in Organic Chemistry with Prof. Karl A. Scheidt from Northwestern University in 2012. She is currently a Marie Curie Post-doctoral Fellow in the group of Prof. Dr. Karl Gademann at the University of Basel (Switzerland), directing projects which explore the isolation, total synthesis, and biological evaluation of neurotogenic natural products.

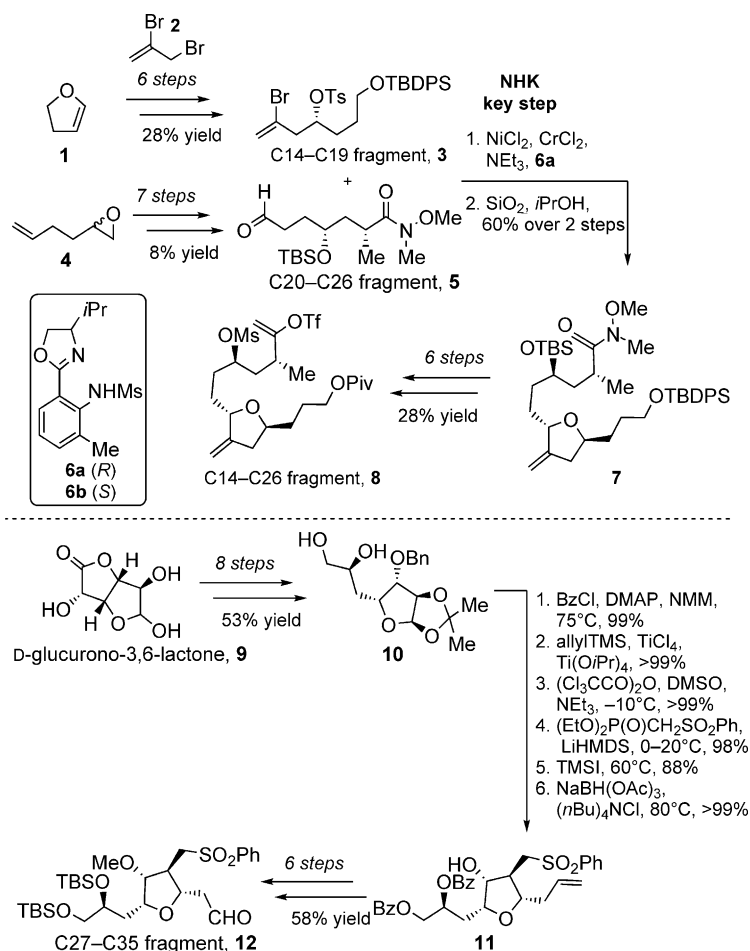


tubulin, and acts as a noncompetitive inhibitor of the binding of radiolabeled vinblastine to tubulin.<sup>[8a,12]</sup>

Despite this phenomenal biological activity, the limited supply (less than  $2 \times 10^{-6}$  wt % yield)<sup>[9b]</sup> of the natural product from its marine source and the potential for contamination with other closely related halichondrins or highly toxic metabolites, such as okadaic acid,<sup>[12]</sup> remained serious obstacles that could only seemingly be circumvented by total synthesis. Furthermore, the amount of material needed to support preclinical development and subsequent clinical trials was estimated to be approximately 10 g.<sup>[8a,13]</sup> If found to be successful, 1–5 kg year<sup>-1</sup> would be necessary to meet commercial needs. Although the initial aquaculture with *Lissodendoryx* sp. seemed promising, the ability of scale-up to satisfy the commercial demands in a reliable manner could not be achieved.<sup>[8a,13]</sup>

Kishi and co-workers reported the first total synthesis of halichondrin B, along with norhalichondrin B, in 1992.<sup>[8a,14]</sup> The route relied on a series of robust Nozaki–Hiyama–Kishi (NHK) reactions, which were used to construct five key bonds in the synthesis.<sup>[8a,14a,d,e,15]</sup> Further improvements were later developed,<sup>[16]</sup> but the optimized synthetic route contained nearly 120 steps. Despite this, the completion of this landmark total synthesis convinced the U.S. National Cancer Institute (NCI) to move halichondrin B into preclinical development in March 1992.<sup>[13c]</sup> The Eisai Research Institute initiated in vitro and in vivo studies of synthetic halichondrin B, along with several analogues provided by the Kishi research group at Harvard University.<sup>[17]</sup> Surprisingly, they found that the C1–C38 macrolide, or the “eastern portion” of the molecule, demonstrated activities within an order of magnitude of the parent compound during an in vivo 3–4 day growth inhibition assay with a DLD-1 human colon cancer cell line (Figure 2).<sup>[8a,18]</sup> They found that the macrolactone could be replaced by a nonhydrolyzable ester bioisostere to prevent susceptibility toward nonspecific esterases. Attempts to truncate further while retaining activity were unsuccessful. For example, the 2,6,9-trioxatricyclo[3.3.2.0]decane was essential for the activity and global conformation of the molecule.<sup>[9b]</sup>

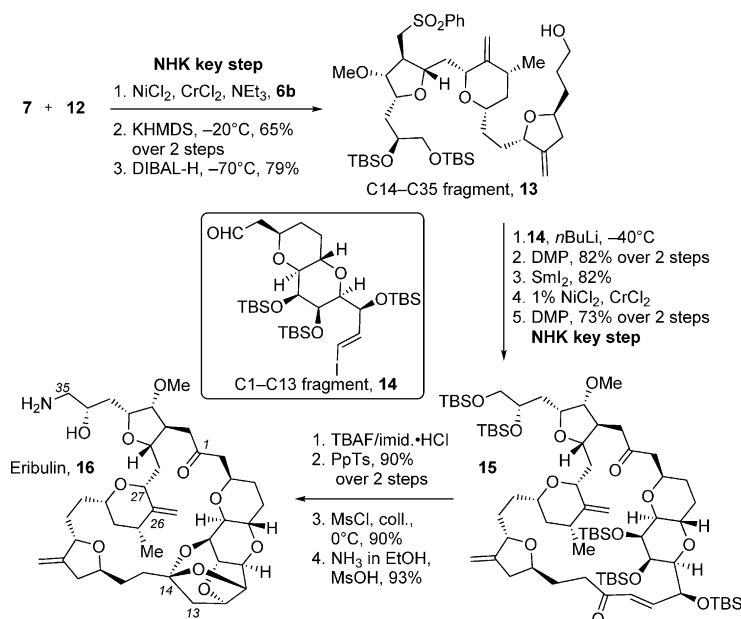
Despite the success with total synthesis, the limited supply of halichondrin B was a critical issue in the early 1990s, which was eased, in particular, by the discovery of the potency of simplified analogue eribulin, or E7389 (Figure 2).<sup>[18a]</sup> In addition to the synthetic efforts by the Kishi group,<sup>[19]</sup> the synthesis of the C14–C35 fragment (**13**) of eribulin (Scheme 2) has been optimized for process development on a kilogram scale. This most recent route put forth by Eisai Inc. in 2013 begins with conversion of dihydrofuran (**1**) into C14–C19 fragment **3**, through a tin-mediated bromoallylation with 2,3-bromopropene (**2**; Scheme 1).<sup>[20]</sup> Fragment C20–C26 (**5**) was synthesized from 1,2-epoxyhex-5-ene (**4**) in seven steps by a hydrolytic kinetic resolution (HKR) with a Jacobsen



**Scheme 1.** Synthesis of the C14–C26 fragment **8** and C27–C35 fragment **12** of eribulin by Kishi and co-workers. Bn = benzyl, Bz = benzoyl, DMAP = 4-dimethylaminopyridine, HMDS = hexamethyldisilazide, Ms = mesyl or methanesulfonyl, NMM = *N*-methylmorpholine, Piv = pivaloyl, TBDPS = *tert*-butyldiphenylsilyl, Tf = trifluoromethanesulfonyl, TMS = trimethylsilyl.

catalyst.<sup>[21]</sup> Aldehyde **5** and fragment **3** were then combined in an asymmetric NHK Ni/Cr coupling with chiral ligand **6a**,<sup>[22]</sup> followed by treatment with silica gel to induce formation of the tetrahydrofuran ring. Elaboration of furan **7** through six steps provided the C14–C26 fragment **8**. Synthesis of the C27–35 fragment **12** commenced with transformation of readily available D-glucurono-3,6-lactone (**9**) into C34–C35 diol **10** in eight steps. Reaction of the tetrahydrofuran ring with allyltrimethylsilane, followed by a Horner–Wadsworth–Emmons olefination and a hydroxy-directed conjugate reduction afforded sulfone **11**. Six additional steps provided the desired C27–35 fragment **12** in a final yield of 26% through an entirely chromatography-free, 20-step process.

The C14–C26 (**8**) and C27–C35 (**12**) subunits were combined in an iterative Nozaki–Hiyama–Kishi reaction and Williamson ether cyclization with ligand **6b** (Scheme 1) in d.r. = 20:1 and a 65 % overall yield. A subsequent reductive cleavage afforded the C14–C35 fragment **13** (Scheme 2).<sup>[23]</sup> Sulfone **13** was then combined with the C1–C13 aldehyde **14** from the halichondrin B synthesis, which was assembled in 13 steps from L-mannonic- $\gamma$ -lactone.<sup>[16b]</sup> A SmI<sub>2</sub>-mediated



**Scheme 2.** Completion of the synthesis of eribulin (**16**). Coll. = 2,4,6-collidine or 2,4,6-trimethylpyridine, DIBAL-H = diisobutylaluminum hydride, DMP = Dess Martin periodinane, Imid. = imidazole, PpTs = pyridinium *para*-toluenesulfonate, TBAF = tetrabutylammonium fluoride, TBS = *tert*-butyldimethylsilyl.

desulfonylation, a key Nozaki–Hiyama–Kishi macrocyclization, followed by oxidation were then performed. The trioxatricyclo[3.3.2.0]decane ring system was installed in macrocycle **15** in 90 % yield by treatment with TBAF and then PpTs. A subsequent transformation of the C35 hydroxy group gave the amine of eribulin (**16**), which was optimized by Kaburagi and Kishi in 2007 to an 84 % overall yield.<sup>[24]</sup> Additionally, the Kishi group reported a special system for the formation of the trioxatricyclo[3.3.2.0]decane ring system, which involved a DOWEX 50WX8-400 sulfonic acid resin and CaCO<sub>3</sub> work-up of the TBAF-mediated TBS cleavage reaction, followed by filtration and evaporation.<sup>[25]</sup> Then, passage of the resulting residue through an ion-exchange resin-based device completed the formation of the ketal<sup>[16c]</sup> in 90 % overall yield.<sup>[26]</sup>

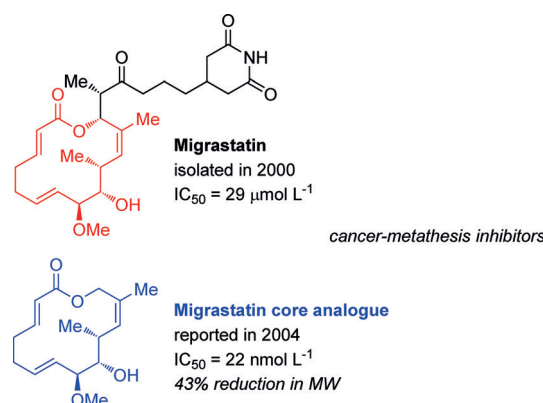
In vivo studies revealed that eribulin possesses excellent activities at 0.1–1 mg kg<sup>-1</sup> against several chemoresistant human solid tumor xenografts.<sup>[18a]</sup> Eribulin displayed superior efficacy over other antimitotics such as even paclitaxel.<sup>[13c]</sup> These studies also confirmed that this analogue could induce G<sub>2</sub>-M cell-cycle arrest and disrupt mitotic spindles, and was consistent with the tubulin-based antimitotic mechanism of the parent halichondrin B. Eisai investigated the safety and effectiveness of eribulin mesylate in a single phase III clinical trial involving 762 women with metastatic breast cancer, during which the median overall survival of patients receiving eribulin mesylate was 13.1 months, whereas this rate was 10.6 months for those receiving only a single agent therapy.<sup>[27]</sup> The U.S. FDA approved eribulin mesylate (Halaven) on November 15, 2010 for patients with metastatic breast cancer, who have received at least two prior chemotherapy treatments for late-stage disease, including anthracycline- and taxane-based therapies. The European Commission has recently followed suit, issuing their approval on July 3, 2014

under the same conditions as the FDA.<sup>[28]</sup> With this approval in the European Union member states, eribulin mesylate is now approved in more than 50 countries worldwide, including Japan, Singapore, and Switzerland. Eisai is currently investigating eribulin mesylate as a treatment for breast cancer with fewer prior treatments, as well as soft-tissue sarcoma and non-small-cell lung cancer.

Overall, the development of eribulin constitutes a vast improvement over the parent compound as it has a large decrease in complexity, including a 35 % reduction in molecular weight, which requires half the number of synthetic steps to access, and retains the potent antitumor activity, as clearly evident by its use today as a cancer treatment agent worldwide.

### 2.1.2. From Migrastatin to a Core Analogue

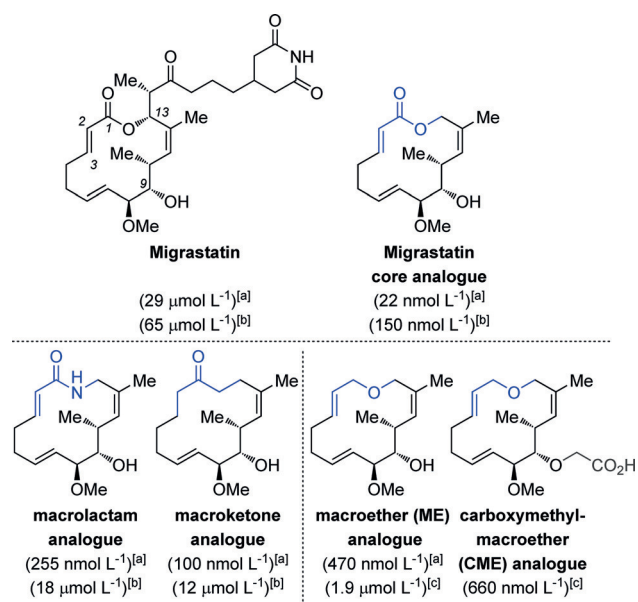
Migrastatin was isolated in 2000 by the Imoto group from a cultured broth of *Streptomyces* sp. MK929-43F1 (Figure 3).<sup>[29]</sup> This 14-membered macrolide with a glutarimide-terminated side chain inhibited the spontaneous migration of EC17 human esophageal and B16 mouse melanoma cells at a concentration of 10–30 µg mL<sup>-1</sup> (IC<sub>50</sub> = 82 µg mL<sup>-1</sup>).<sup>[29c]</sup>



**Figure 3.** Structural comparison between migrastatin and the migrastatin core analogue. IC<sub>50</sub> values are for 4T1 murine breast cancer.

However, the compound demonstrated no appreciable cytotoxicity, inhibition of protein synthesis, or antibiotic/antifungal activities. In a follow-up study, the ability of migrastatin to inhibit the growth of Ms-1 human small cell lung carcinoma cells under anchorage-independent conditions in a dose-dependent manner (1–100 µg mL<sup>-1</sup>) implied its influence on integrin signaling, which is also involved in cell migration.<sup>[29c]</sup> A year after the absolute configuration was confirmed by X-ray crystallography,<sup>[30]</sup> the Danishefsky group reported the first synthesis of the macrolide by utilizing a Lewis acid catalyzed diene aldehyde condensation (LACDAC) and ring-closing metathesis (RCM).<sup>[31]</sup>

Follow-up SAR investigations by the Danishefsky group revealed that, out of eight analogues screened, the migrastatin

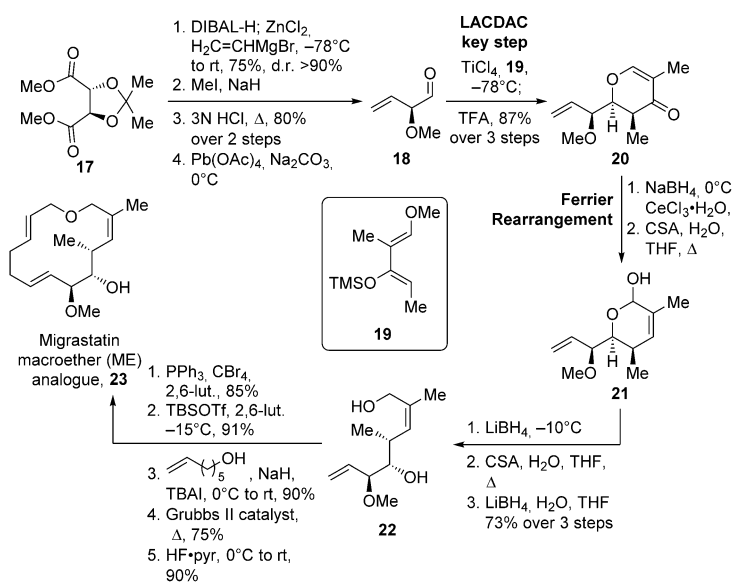


**Figure 4.** Comparison of the structures and activities of migrastatin analogues from chamber cell migration assays.  $\text{IC}_{50}$  values in parentheses are with the following cell lines: [a] 4T1 murine breast cancer,<sup>[31c,32]</sup> [b] HUVEC human umbilical vein endothelial cells,<sup>[31c]</sup> and [c] A549 human lung cancer.<sup>[33]</sup>

core surpassed the activity of the parent compound in a Boyden chamber cell migration assay with serum-induced 4T1 mouse breast tumor cells by three orders of magnitude, with an  $\text{IC}_{50}$  value of 22  $\text{nmol L}^{-1}$ , versus the 29  $\mu\text{mol L}^{-1}$  value for migrastatin (Figure 4).<sup>[32a]</sup> Analysis of other analogues revealed that reduction of the C2–C3 double bond could be tolerated, while modification of the hydroxy group at C9 or at C13, or at the ketone at C1 could not be tolerated with respect to the potent cell migration suppression activity.<sup>[31c]</sup> Eight analogues and migrastatin were also tested for their metabolic stability in mouse plasma and, while the parent compound and compounds containing a glutarimide side chain were inert, the most active compounds including the migrastatin core analogue were hydrolyzed rapidly by esterases. These findings resulted in the macrolactam and macroketone analogues being lead compounds from these studies, with  $\text{IC}_{50}$  values of 255  $\text{nmol L}^{-1}$  and 100  $\text{nmol L}^{-1}$ , respectively, and with no esterase susceptibility.<sup>[34a]</sup> As it was suspected that these analogues influence the angiogenesis process, Danishefsky and co-workers also investigated endothelial cell migration with human umbilical vein endothelial cells (HUVECs). These results mirrored the previous ones, in that the analogues showed more activity than the parent compound, albeit with some erosion of potency, and they found that the macrolactam, macroketone, and the macrolactone analogues did not have any cytotoxic or antiproliferative effects up to 20  $\mu\text{mol L}^{-1}$  in an in vitro mouse assay.<sup>[32a]</sup> This observation confirmed that inhibition of cell proliferation is not a contrib-

utor to the effects observed in the chamber assays. A year later in 2005, Danishefsky and co-workers reported that several analogues, including the migrastatin macrolactam analogue, inhibit mammary tumor metastasis in mice, which suggests that these compounds interfere with the invasion step by demonstrating that they block Rac activation, lamellipodia formation, and cell migration.<sup>[34]</sup>

In a 2010 report, the Danishefsky group reported an even more simple, new macroether (ME) analogue **23** that retains the cancer cell migration inhibition activity of the analogue family (Scheme 3).<sup>[32b]</sup> The synthetic sequence towards ME analogue **23** shared the first nine steps of the original route towards the natural product.<sup>[31a–c]</sup> The synthesis began with transformation of tartrate derivative **17** into aldehyde **18** by utilizing a highly diastereoselective divinylzinc addition. The key LACDAC reaction of aldehyde **18** with diene **19** was performed under  $\alpha$ -chelation control in the presence of the Lewis acid  $\text{TiCl}_4$ , and then cyclization with TFA proceeded in 87% yield to afford dihydropyranone **20**. Selective 1,2-reduction of the ketone and treatment with CSA promoted a Ferrier rearrangement to the corresponding lactol **21**, which was then transformed into primary alcohol **22** in three steps. A Williamson etherification reaction and RCM afforded ME analogue **23** in 47% yield over the five steps. Overall, this



**Scheme 3.** Synthesis of the migrastatin macroether analogue by Danishefsky and co-workers (ME, **23**). CSA = camphorsulfonic acid, DCM = dichloromethane, 2,6-Lut. = 2,6-lutidine, pyr = pyridine, TBAI = tetrabutylammonium iodide, TFA = trifluoroacetic acid, TBSOTf = *tert*-butyldimethylsilyltrifluoromethane sulfonate.

analogue had a significant reduction in the complexity of migrastatin, yet retained the activity of the parent compound, and was accessed in 11 fewer synthetic steps.

In cell-migration assays, ME analogue **23** demonstrated migration inhibition against several breast cancer cell lines, including the highly metastatic human LM2, without affecting



the viability of the cells.<sup>[32b]</sup> This analogue also inhibited tumor invasion and metathesis, and prolonged the overall survival in NOD/SCID mice injected with human breast cancer MDA-MD-231 cells in the abdominal mammary fat pad (human breast cancer xenograft model).<sup>[33]</sup> In the course of these studies, the authors also proposed that **23** interferes with fascin-1-dependent migratory behavior. This interaction was confirmed by Chen et al. in 2010 with the X-ray crystal structure of the macroketone analogue in actin-binding sites of fascin.<sup>[35]</sup> However, there was some debate over this result, as the X-ray structure of the macroketone analogue contained an *E*- instead of the *Z*-configured double bond between C4 and C5, and the C6 stereo-center was inverted to the *S* configuration.<sup>[36]</sup>

In 2011, Danishefsky and co-workers reported that **23** was considered to be the most promising analogue on the basis of its *in vitro* and *in vivo* activities, physical properties, ease of preparation, and biological stability (Figure 4).<sup>[33]</sup> This publication details the promising *in vitro* and *in vivo* migration inhibition activity of ME against metastatic small-cell lung carcinoma (SCLC), and also introduced yet another promising candidate, carboxymethyl-ME (CME), which was synthesized with an aim to enhance the bioavailability and pharmacostability, and achieved a higher potency with lower toxicity than ME. This CME analogue was also used as evidence against the X-ray crystallography studies of Chen et al.: since the carboxylic acid functionality reduced cell membrane permeability, and this compound retained its potent metastasis inhibition, the findings implied that the primary target is not an intracellular protein such as Fascin, but a protein target on the cell surface. In 2013, Majchrzak et al. added to this debate by reporting that the migrastatin core analogue showed antibranching activity and interfered with the mechanisms of filopodia assembly, thereby preventing cross-linking of fascin-1-dependent actin filaments *in vitro*.<sup>[37]</sup> Overexpression of *fascin1* in cancer cells has been linked with clinically aggressive tumors, poor prognosis, and shortened disease-free survival. Further studies on these migrastatin analogues and additional structures are ongoing.

### 2.1.3. From Anguinomycin C/D to an Anguinomycin Analogue

In 1995, Hayakawa et al. reported the structures of anguinomycins C and D, which were isolated from cultures of *Streptomyces* sp. (Figure 5).<sup>[38]</sup> These antitumor antibiotics induced cell death in pRB-inactivated glia cells at picomolar concentrations and, surprisingly, cell-cycle arrest at only the G1 phase was observed for normal rat glia cells. Retinoblastoma tumor suppressor protein (pRB) is often inactivated during the development of a variety of cancers.

The absolute configuration of anguinomycin C was confirmed by the first total synthesis in 2007 by Gademann and co-workers. This route had a total of 29 steps, with 18 steps in the longest linear sequence, and an overall yield of 6.7%.<sup>[39]</sup> Biological studies demonstrated that this natural product is a potent inhibitor of CRM1-dependent protein export from the cell nucleus, an activity shared by a close structural

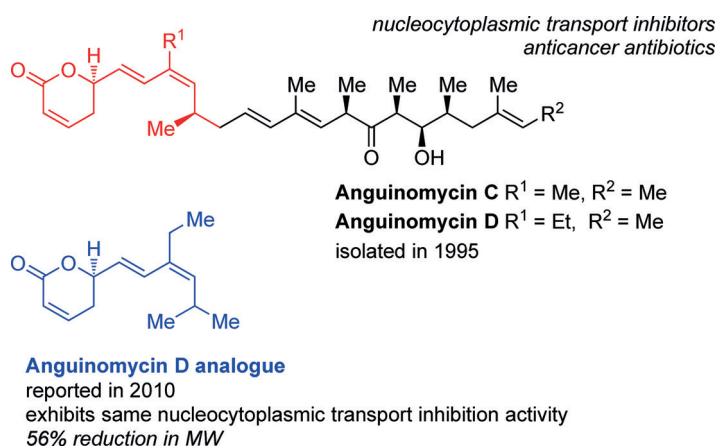
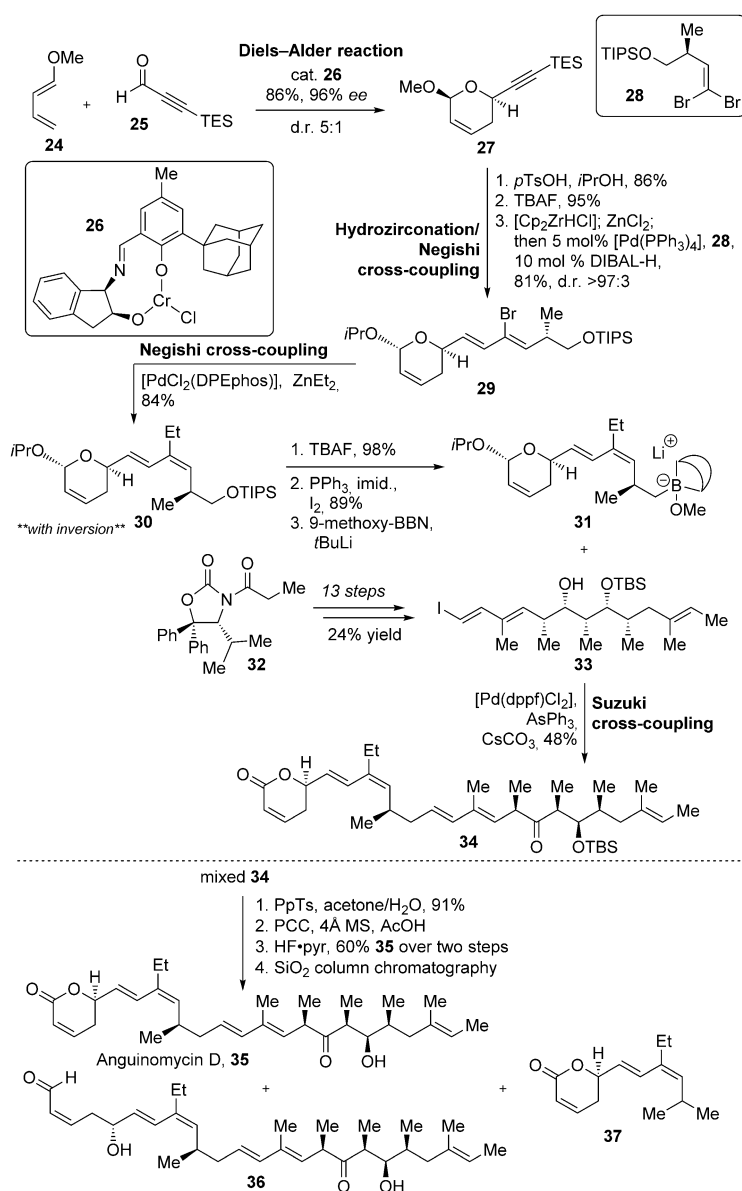


Figure 5. Structural comparison between anguinomycin C/D and the anguinomycin D analogue.

relative, leptomycin B. It is known that leptomycin B covalently modifies chromosomal region maintenance 1 (CRM1; exportin 1) at the nucleophilic sulphhydryl group of Cys528 by utilizing its  $\alpha,\beta$ -unsaturated  $\delta$ -lactone, thus preventing export of protein cargoes that rely on this cleft by preventing formation of the ternary CRM1/cargo substrate/Ran complex, or the binary complex CRM1/cargo substrate in the absence of Ran.<sup>[40]</sup> More recent computational studies of inhibitors, such as leptomycin B, to CRM1 revealed that the mechanism of inhibition goes beyond simple Michael addition, and is followed by a CRM1-mediated hydrolysis of the lactone.<sup>[41]</sup> Mutagenesis revealed that at least one of the residues Arg543, Lys548, or Cys579 needs to be present to stabilize the resulting anionic tetrahedral intermediate and lower the energy barrier to drive the reaction. Furthermore, evidence not only supported that hydrolysis must follow conjugate addition, as the corresponding hydroxy acids do not inhibit CRM1, but also that hydrolysis decreases the reversibility of the Michael addition and enables persistent binding of the inhibitors to CRM1. This argument is supported by the fact that reversibility of the conjugate addition should be kinetically controlled and the  $\alpha$ -proton of the hydrolyzed (carboxylate) intermediate is much less acidic than the corresponding proton of the lactone.<sup>[41]</sup>

Although dose-limiting toxicity prevented further development of leptomycin B as an antitumor agent,<sup>[42]</sup> Gademann and co-workers saw the potential of the anguinomycin core and went on to synthesize anguinomycin D and analogues for further studies with the aim of investigating if the polyketide side chain mimics the hydrophobic leucine-rich nuclear export signal of the cargo protein and is necessary for activity.<sup>[43]</sup> The route employed was similar to the synthesis of the anguinomycin C derivative, and began with an asymmetric hetero-Diels–Alder reaction between diene **24** and aldehyde **25** with catalyst **26** according to Jacobsen et al.,<sup>[44]</sup> which afforded pyran **27** in 86% yield and with 96% *ee* (Scheme 4). The acetal was inverted to the more thermodynamically stable compound and a hydrozirconation of the alkyne with the Schwartz reagent, followed by transmetalation with Zn for a Negishi cross-coupling utilizing  $[\text{PdCl}_2$ -



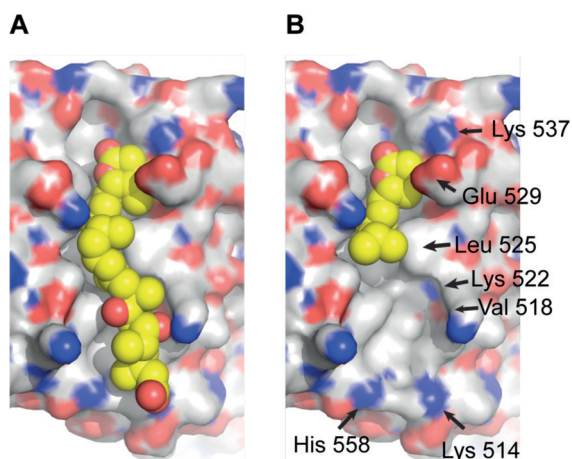
(DPEphos)] with dibromoolefin **28**<sup>[45]</sup> to afford diene **29**, but with inversion to the undesired isomer. Inversion to the desired *Z* olefin was accomplished with another Negishi cross-coupling reaction to install the ethyl group in 84% yield and with good selectivity. Silyl ether **30** was then converted into boronate **31**. A classic, Evans *syn*-aldol strategy was employed to assemble vinyl iodide **33** with the DIOZ (4-isopropyl-5,5-diphenyloxazolidin-2-one) auxiliary **32** developed by Hintermann and Seebach.<sup>[46]</sup> The 13 step route, which was executed in 24% overall yield, employed the DIOZ auxiliary three times and two boron-mediated aldol reactions. Boronate **31** and vinyl iodide **33** were combined under Suzuki coupling conditions developed by Marshall

et al.<sup>[47]</sup> With a particular batch of the Suzuki reaction, there was a minor by-product, which was taken with desired product **34** through the last three steps and purified by column chromatography to afford not only anguinomycin D (**35**), but also aldehyde analogue **36** and truncated analogue **37**, likely a degradation product of the boronate intermediate from the previous Suzuki coupling.

These compounds were then screened for their ability to inhibit CRM1-dependent protein export by measuring the accumulation of Rio2 protein in the HeLa cell nucleus by indirect immunofluorescence analysis. Anguinomycin C and D (Figure 5) both displayed partial inhibition at 5 nmol L<sup>-1</sup> and full inhibition at 10 nmol L<sup>-1</sup> while, surprisingly, aldehyde analogue **36** showed full inhibition at 50 nmol L<sup>-1</sup> and truncated analogue **37** exhibited an unexpected strong inhibition at 25 nmol L<sup>-1</sup>. This study demonstrated that the activity could be retained even when the anguinomycin polyketide chain was replaced by a hydrophobic moiety or when completely removed, and reinforced the importance of the conjugate acceptor ability of the  $\alpha,\beta$ -unsaturated  $\delta$ -lactone.<sup>[41]</sup> This concept that the anguinomycin core could be reduced by nearly 60% in molecular weight and still retain its activity was supported by molecular modeling studies of the (*R*)- $\alpha,\beta$ -unsaturated  $\delta$ -lactone in the NES (nuclear export signals) binding pocket of CRM1<sup>[40c-e]</sup> with all-atom energy minimization (Figure 6).<sup>[49]</sup> Further investigations into this more synthetically accessible, truncated analogue **37** of anguinomycin could provide more powerful or selective nucleocytoplasmic transport inhibitors for cancer treatment.<sup>[50]</sup>

#### 2.1.4. From Duocarmycin SA to N-Boc-DSA

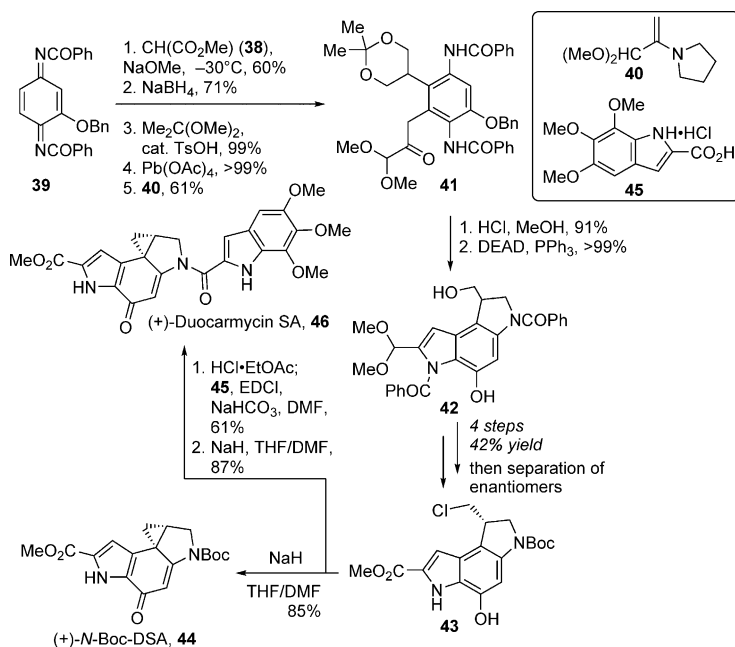
Duocarmycin SA was isolated from *Streptomyces* sp. DO113 collected from a soil sample at the Rokkakudo temple in Kyoto, Japan in 1990 by Takahashi and co-workers, and given its name because it was more stable and demonstrated more potent Gram-positive antibacterial (minimum inhibition concentrations (MICs): 2.7 nmol L<sup>-1</sup> for *Staphylococcus aureus* and 1.4 nmol L<sup>-1</sup> for *Bacillus subtilis*) and cytotoxic activities (against murine lymphocytic leukemia P388 and murine sarcoma 180) than close relative duocarmycin A (Figure 7).<sup>[51]</sup> The duocarmycins act as sequence-selective DNA alkylating agents with 3'→5' binding directionality, specific for AT-rich regions in the minor groove, where stabilizing van der Waals contacts are maximized.<sup>[52]</sup> Binding of the agent to DNA induces a conformational change, which disrupts the stabilization of the vinylogous amide and activates the cyclopropane ring of the cyclopropapyrroloindole in the left-hand side of the core for the reversible, stereoelectronically controlled adenine-N3 addition in an S<sub>N</sub>2 fashion (shape-dependent catalysis).<sup>[52b]</sup> The bound agent spans 3.5 base pairs and complements the topological curvature and pitch of the minor groove, with the



**Figure 6.** Computational model of A) leptomycin B and B) truncated analogue **37** binding to CRM1 (C: gray, O: red, N: blue; C in inhibitor molecule: yellow).<sup>[48]</sup>

hydrophobic face of the molecule implanted deeply within.<sup>[53]</sup> SAR studies of another close relative, (+)-CC-1065, identified the required, but not necessarily optimal, pharmacophore for this family of molecules, thus rendering further studies on the more active and stable duocarmycin SA necessary.

The first total synthesis of duocarmycin SA (**46**), as well as the synthesis of analogue **44** was reported in 1992 by Boger and Machiya (Scheme 5).<sup>[54]</sup> Two successive regioselective alkylations of substituted diimide quinone **39** (synthesized in 5 steps and 38 % yield)<sup>[55]</sup> were performed at C5 with dimethyl malonate (**38**), and then at C6 with pyrrolidine enamine of pyruvaldehyde dimethyl acetal **40**<sup>[56]</sup> to afford aniline **41**. Acidic conditions removed the acetal group and promoted



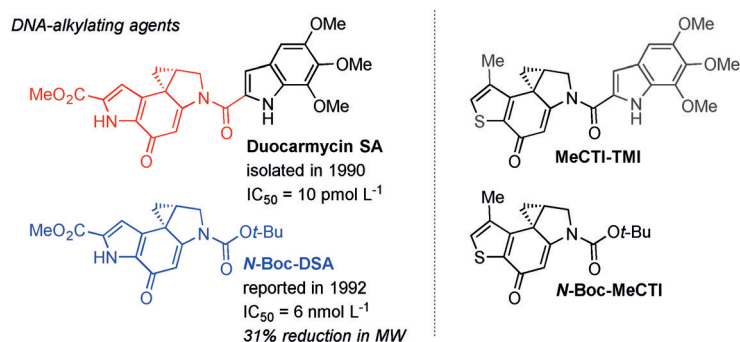
**Scheme 5.** Total synthesis of (+)-duocarmycin SA (**46**) and (+)-*N*-Boc-DSA (**44**) by Machiya and Boger. DEAD = diethyl azodicarboxylate, EDCI = 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide, DSA = duocarmycin SA.

indole formation, which was followed by cyclization of the resulting diol to complete the dihydropyrroloindole core **42**. Eight additional steps, including chiral resolution of the bis-(*R*)-*O*-acetylmandelate esters, provided (+)-dihydropyrroloindole **43**. This intermediate could then be directly transformed into (+)-*N*-Boc-DSA analogue **44** in 85 % yield, or to the natural antipode of (+)-duocarmycin SA (**46**) by coupling with the 5,6,7-trimethoxyindole-2-carboxylic acid salt **45** (made in 3 steps and 74 % yield from 3,4,5-trimethoxybenzaldehyde)<sup>[53]</sup> in 53 % yield over the two steps. The unnatural (–)-*N*-Boc-DSA and (–)-duocarmycin SA enantiomers were also synthesized from (–)-*ent*-**43**.

The natural (+)-enantiomers were found to have 10 times more potent in vitro cytotoxicity than the corresponding unnatural (–) enantiomers, with IC<sub>50</sub> values (L1210 mouse lymphocytic leukemia) of 10 pmol L<sup>–1</sup> for (+)-duocarmycin SA and 6 nmol L<sup>–1</sup> for (+)-*N*-Boc-DSA.<sup>[54b]</sup> *N*-Boc-DSA was found to be a substantially less efficient (ca. 10<sup>4</sup> times) DNA alkylator, with a less strict selectivity profile than the parent natural product (SA), and with both enantiomers showing the same site reactivity profile.<sup>[52a]</sup> The latter property was also unusual, but a natural consequence of the diastereomeric relationship of the adducts. *N*-Boc-DSA was found to be the most stable of the duocarmycin analogues, as measured by chemical solvolysis, with complete stability at pH 7 and a half-life of 177 h at pH 3.<sup>[54b]</sup> Gas-phase heats of reaction for the alkylation of *N*-methyladenine with *N*-acetyl duocarmycin A and *N*-acetyl duocarmycin SA were calculated by using semiempirical methods (AM1, MNDO). The results reflected the experimental observations that the alkylation of DNA is reversible (unlike with (+)-CC-1065), as the model calculations with methyladenine gave a near thermally neutral reaction.<sup>[54b]</sup> This underlines the importance of noncovalent interactions in the minor groove (hydrophobic binding/van der Waals contacts), as DNA is nonetheless effectively alkylated by duocarmycins (binding-driven bonding).

A fundamental parabolic relationship for all the duocarmycins was established between the reactivity of the electrophilic cyclopropane ring and the cytotoxic activity spanning a 10<sup>4</sup>–10<sup>6</sup> range, where an increased solvolytic stability or decreased reactivity correlates to an increased cytotoxic potency.<sup>[54b,57]</sup> Highly stable analogues do not efficiently alkylate the DNA, while highly reactive analogues do not reach the biological target. For proof of concept, Boger and co-workers introduced the rationally designed thiophene analogue *N*-Boc-MeCTI in 2007, which reflected the optimal point of balanced stability and reactivity in the parabolic relationship, being slightly more stable, but 5–6 times less potent than *N*-Boc-DSA (Figure 7).<sup>[57,58]</sup> In this study, they also reported analogue MeCTI-TMI, which has an even higher cytotoxic potency than duocarmycin SA (IC<sub>50</sub> = 5 pmol L<sup>–1</sup> versus 10 pmol L<sup>–1</sup>). The retained single-digit nanomolar potency of *N*-Boc-DSA despite a significant reduction in structural complexity rendered it an excellent example of capturing biological activity with a natural product fragment.

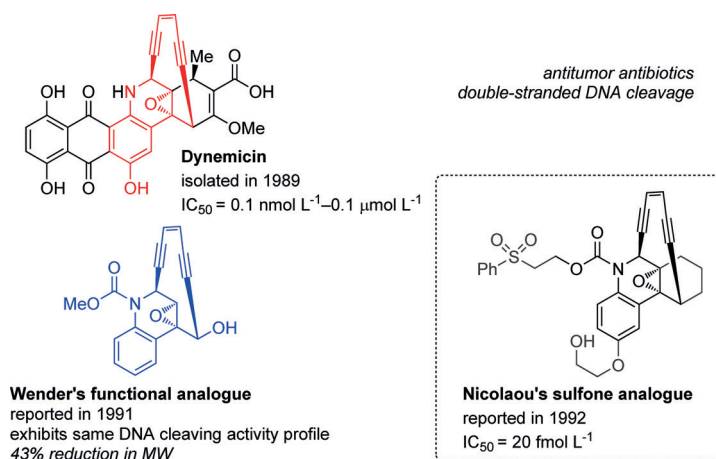




**Figure 7.** Structural comparison of duocarmycin SA and analogues.  $IC_{50}$  values are for L1210 mouse lymphocytic leukemia. Boc = *tert*-butoxycarbonyl, MeCTI = 7-methyl-1,2,8,8a-tetrahydrocyclopropa[*c*]thieno[3,2-*e*]indol-4-one, TMI = 5,6,7-trimethoxyindole-2-carboxylate.

### 2.1.5. From Dynemicin A to a Dynemicin Analogue

The violet dynemicin A was discovered in 1989 in the fermentation broth of *Micromonospora chersina* sp. (M956-1), isolated from a soil sample from Gujarat State, India (Figure 8).<sup>[59]</sup> It demonstrated potent Gram-positive antibacterial activity with low toxicity, especially against *Staphylococcus aureus* Smith infection (mouse i.p.,  $PD_{50}$  =



**Figure 8.** Structural comparison of dynemicin A and analogues.  $IC_{50}$  values are for Molt-4 T-cell leukemia.

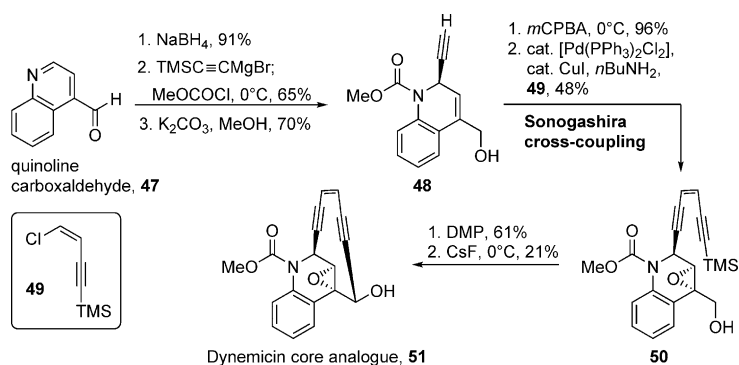
$0.13 \text{ mg kg}^{-1}$ ), and potent in vivo antitumor activity against B16 melanoma, Moser human carcinoma, HCT-116 human carcinoma, and normal/vincristine-resistant P388 leukemia ( $IC_{50}$  = 7–9  $\text{nmol L}^{-1}$ ), even prolonging the life span of mice with P388 leukemia or B16 melanoma. Structurally, it contains a unique hybrid of an anthraquinone and a 1,5-diyne-3-ene system embedded within a strained 10-membered ring. The enediyne unit makes it a relative of the esperamicin/calicheamicin family. The mechanism of action involves intercalation of the anthraquinone core of into the minor groove of the DNA helix, followed by attack of the phenyl diradical resulting from the enediyne core, thereby leading to DNA strand scission of the sugar–phosphate backbone three

base-pairs apart.<sup>[60]</sup> The phenyl diradical is proposed to form by two sequential one-electron reductions of the hydroquinone to the hydroquinonediol, epoxide opening (key to activation), nucleophilic attack (i.e. water) on the *p*-quinone methide intermediate or simple tautomerization, and then Bergman cyclization of the strained (*Z*)-enediyne to the diaryl radical. Dynemicin A preferentially cuts the 3' side of purine bases with a preference for guanines, thus differing from its esperamicin/calicheamicin relatives, and demonstrates a preference for double-stranded (B-form) and stem regions of single-stranded DNA.

Along with several pioneering synthetic studies,<sup>[61]</sup> the first total synthesis and absolute configuration of (+)-dynemicin A was reported by Myers et al. in 1995.<sup>[62]</sup> In total, this route was completed in 26 steps in 0.3 % yield and employed an *exo*-selective Diels–Alder reaction as the key step. Several analogues were also made by using the same approach. In the following years, copious synthetic and SAR studies of enediyne analogues of the dynemicin core were performed by the groups of Schreiber,<sup>[61a,d]</sup> Wender,<sup>[63]</sup> Nicolaou,<sup>[61b,64]</sup> Isobe,<sup>[65]</sup> Myers,<sup>[62a,66]</sup> Danishefsky,<sup>[61g,67]</sup> Maier,<sup>[68]</sup> Magnus,<sup>[69]</sup> and others.<sup>[64a,d,70]</sup> These studies probed the effects of triggering groups or initiators on the nitrogen atom or aryl ring, which could be activated under basic or photochemical conditions that could be mimicked by intracellular processes. Tethering devices were also investigated to aid target delivery, as were deactivating groups that would modulate enediyne activity, as well as detection devices that would facilitate mechanistic studies.<sup>[64a]</sup> One of the most potent dynemicin analogues was the sulfone analogue of Nicolaou et al. with an  $IC_{50}$  value of 20 fm against Molt-4 T-cell leukemia, as compared to an  $IC_{50}$  range of  $0.1 \text{ nmol L}^{-1}$ – $0.1 \text{ } \mu\text{mol L}^{-1}$  for dynemicin A (Figure 8).<sup>[64a,b,d,71]</sup> However, one of the most simplified analogues was accessed by Wender et al. in just 7 steps (Scheme 6).<sup>[63a]</sup> The route began with the reduction of commercially available quinoline carboxyaldehyde (**47**) and then stepwise incorporation of the enediyne bridge, first by ethynyl Grignard addition to the imine to give alkyne **48**, followed by formation of the key epoxide and Sonogashira coupling with enyne **49**. Oxidation of alcohol **50** to the aldehyde and treatment with cesium fluoride permitted closure of the strained 10-membered ring, thereby producing analogue **51**. As compared to the parent compound, this greatly simplified analogue exhibited the same activity profile for cleaving plasmid DNA.<sup>[63b,d]</sup>

### 2.1.6. From Bryostatin 1 to Picolog

The bryostatin family consists of 20 complex natural products that were isolated from the marine bryozoan *Bulgula neritina*, starting in the late 1960s.<sup>[72]</sup> They all share a 20-membered macrolide core with three densely substituted tetrahydropyran rings and a signature exocyclic enoate appended to at least one of these rings. The actual source of these molecules has been narrowed down to a bacterial



**Scheme 6.** Synthesis of the dynemicin core analogue (**51**) by Wender and co-workers. mCPBA = *meta*-chloroperbenzoic acid.

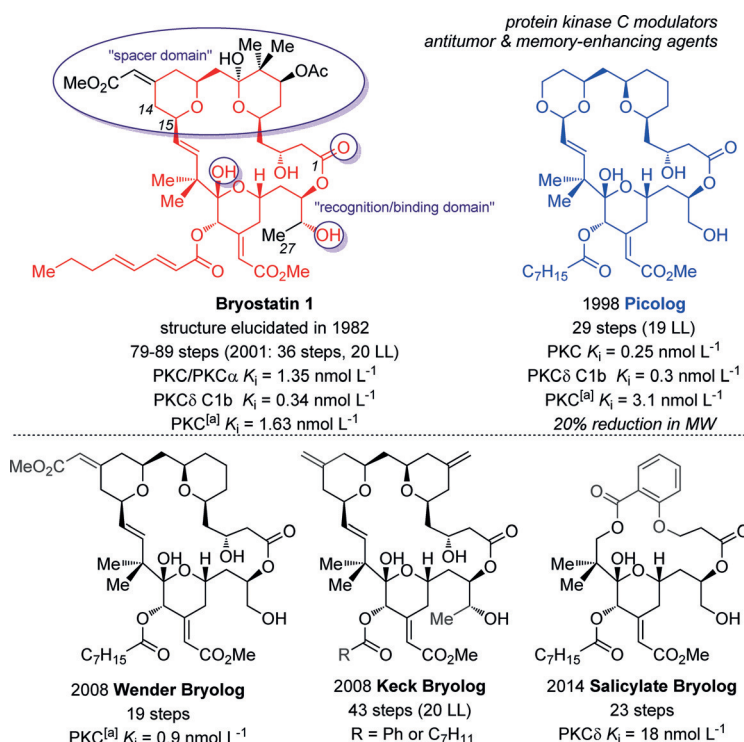
symbiont of *B. neritina*, *Endobugula sertula*, which uses them to protect its larvae from predators.<sup>[73]</sup> The most well-studied member, bryostatin 1, has a diverse and extremely important range of biological activities stemming from its ability to modulate protein kinase C (PKC; Figure 9).<sup>[74]</sup> Bryostatin 1 exhibits potent anticancer activity by reversing drug resist-

clinical trials for Alzheimer's disease, with another trial currently ongoing. In addition, bryostatin 1 is also currently being evaluated for HIV/AIDS treatment in yet another clinical trial. A recent study confirmed its ability to activate latent viral reservoirs and downregulate the CD4 cell surface receptor synthesis/expression necessary for viral entry into uninfected cells.<sup>[78]</sup> Other potential therapeutic applications include treatments for cardiovascular disease, stroke, pain, or cognitive dysfunction.

Despite the remarkable biological activity of the bryostatins, the clinical supply by good manufacturing practice (GMP) is dwindling because of the natural scarcity (isolation yields of  $1 \times 10^{-3}$  to  $1 \times 10^{-8}$  %), the lack of sustainable success with aquaculture or biosynthesis, and the lack of facile or scalable routes to these molecules, notwithstanding the synthetic advances of the last 30 years. Currently, seven total syntheses of bryostatins 7, 2, 3, 16, 1, and 9 have been reported by the groups of Masamune,<sup>[79]</sup> Evans,<sup>[80]</sup> Yamamura,<sup>[81]</sup> Trost,<sup>[82]</sup> Keck,<sup>[83]</sup> Wender,<sup>[84]</sup> and Krische.<sup>[85]</sup> The initial routes were completed in 79–89 steps, with the most recent route towards bryostatin 7 by Krische and co-workers in 2011 being the most efficient with a total step count of 36 (longest linear sequence: 20 steps).

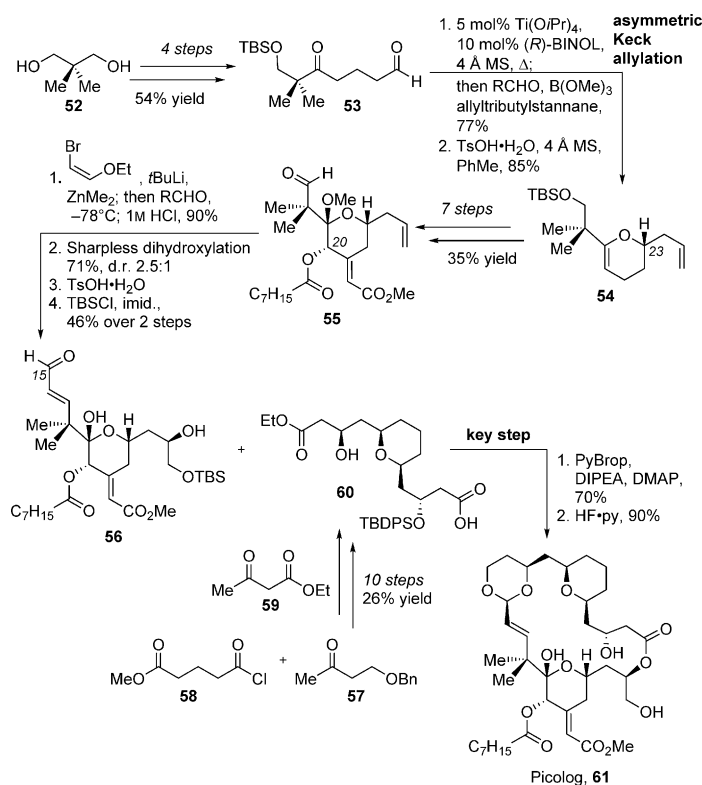
In an effort to avoid stepwise limitations of total synthesis, and the dose-limiting toxicities of the parent compound, extensive studies on the synthesis and evaluation of bryostatin analogues (bryologs) have been performed, most comprehensively by the Wender research group since the 1980s. To date, over 100 analogues have been synthesized and screened for various activities, and nearly one-third exhibit single-digit nanomolar or even picomolar potencies for PKC binding.<sup>[6d]</sup> These studies were primarily driven by the guiding principle that the bryostatin core structure could be divided into the upper "binding" or "recognition domain" (C1–C14) and lower "spacer domain" (C15–C27) according to its interactions with PKC (Figure 9).<sup>[86]</sup> Studies on bryologs have examined the significance of the A ring,<sup>[87]</sup> the B ring,<sup>[88]</sup> the C20<sup>[89]</sup> and C7 side chains,<sup>[90]</sup> as well as de novo structures,<sup>[91]</sup> among others.<sup>[92]</sup> Notable examples of analogues include picolog synthesized by Wender et al. (reported in 2002 with a PKC binding affinity of  $0.25 \text{ nmol L}^{-1}$ ), which was synthesized in 29 steps with a longest linear sequence of 19 steps.<sup>[93]</sup> This represented one of the most potent and promising simplified bryologs, as this was realized in 50 fewer synthetic steps than the syntheses towards the parent structure at that time, and surpassed its activity (bryostatin 1, PKC  $K_i = 1.35 \text{ nmol L}^{-1}$ ).

The synthesis of picolog (**61**) began with the conversion of diol **52** into aldehyde **53** in four steps so that an asymmetric Keck allylation could set the stereocenter at C23 and an acid-catalyzed cyclization/dehydration could form lactone **54** (Scheme 7).<sup>[93b,c]</sup> An additional, straightforward seven steps elaborated the pyran ring, including installation of the acyl



**Figure 9.** Structural comparison of bryostatin 1 to bryologs (LL = longest linear sequence). [a] Protein kinase C rat brain mix.

ance, stimulating the immune system, or restoring apoptotic function.<sup>[75]</sup> It either has been or is currently being evaluated in 37 clinical trials for various types of cancer.<sup>[76]</sup> It has also been shown to enhance memory and learning in animals, reduce Aβ peptide build-up in mice, induce synaptic contact formation, and reduce postischemic/hypoxic damage resulting from stroke.<sup>[77]</sup> As such, it has been evaluated in two



**Scheme 7.** Synthesis of picolog (**61**) by Wender and co-workers. DIPEA = diisopropylethylamine, PyBrop = bromotripyrrolidinophosphonium hexafluorophosphate.

octanoic acid side chain at C20, to give aldehyde **55**. The enal was installed at C15 through a zinc-mediated addition of (*Z*)-bromo-2-ethoxyethene, followed by acid-induced elimination and then Sharpless dihydroxylation, cleavage of the pyran ketal, and protection with a silyl group gave intermediate **56**, precursor to the recognition domain. The spacer domain **60** was synthesized in 10 steps from 4-benzyloxy-2-butanone (**57**) and methyl 5-chloro-5-oxovalerate (**58**) by employing two asymmetric Noyori hydrogenations and a dienolate addition of ethyl acetoacetate (**59**) in between. The two domains were combined by utilizing a PyBrop-mediated esterification and a deprotection/macro-transacetalization step to afford picolog (**61**) in a 63% yield over the two steps.

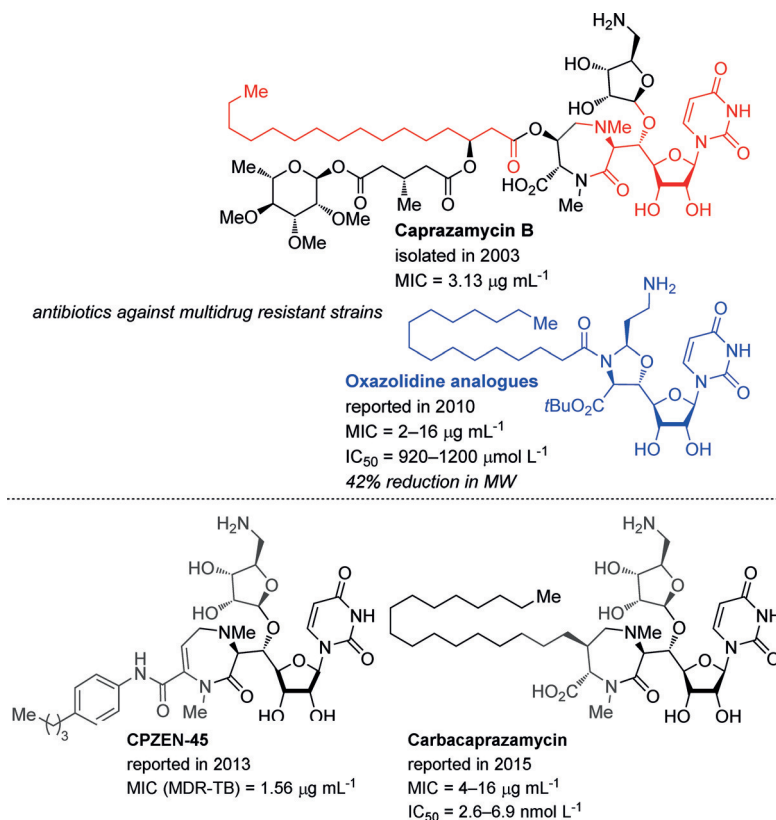
As the original synthetic routes towards the parent bryostatin compounds were limited by a Julia olefination/lactonization sequence, the groups of both Wender and Keck sought a new approach and developed a Prins macrocyclization<sup>[94]</sup> as a key step in back-to-back reports in 2008 (Figure 9). This novel tactic allowed access to more complex bryologs in fewer steps than the parent compounds and with even higher potency than picolog.<sup>[95]</sup> This same powerful strategy was used by Wender and co-workers in 2012 to access bryologs that demonstrated 1000-fold higher

potency than prostratin, the leading clinical candidate at the time.<sup>[78b]</sup> More recent progress has culminated in the Merle series of bryologs from Keck and co-workers,<sup>[96]</sup> and the most simplified bryologs to-date from Wender et al.—the salicylate-derived class (Figure 9).<sup>[97]</sup> This bryolog shown can be accessed in only 23 steps and displays nanomolar PKC binding affinities. The evolution of the bryolog structures and their broad therapeutic applications has been advancing now for more than 40 years and continues to provide a source of inspiration for discovery.

## 2.2. Antibiotic Activity

### 2.2.1. From Caprazamycin B to an Oxazolidinone analogue

Caprazamycin B (CPZ-B) was isolated in 2003 by Igarashi et al. from the culture broth of the actinomycete, *Streptomyces* sp. MK730-62F2 (Figure 10).<sup>[98]</sup> This antimicrobial agent demonstrated excellent activity against drug-susceptible and multidrug-resistant *M. tuberculosis* (MDR-TB or XDR-TB), without significant toxicity in mice. The caprazamycin family (CPZs) are part of the 6'-*N*-alkyl-5'- $\beta$ -*O*-aminoribosyl-glycluridine class of antibiotics, and include liposidomycins (LPMs), which exhibit excellent antimicrobial activities against Gram-positive bacteria.<sup>[99]</sup> The mode of action of these liponucleosides is inhibition of peptidoglycan synthesis by



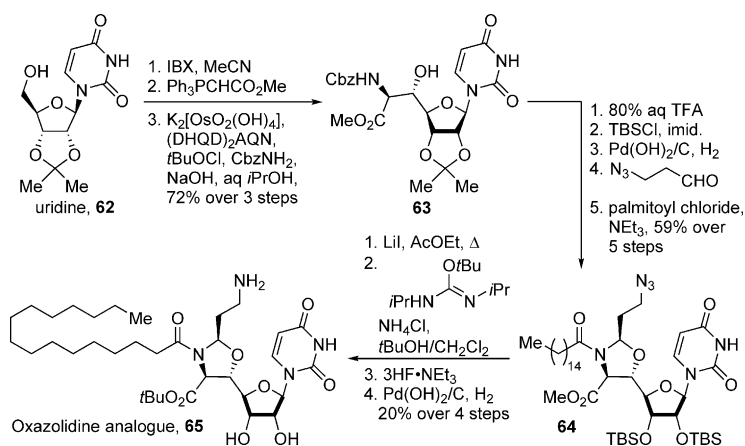
**Figure 10.** Structural comparison of caprazamycin B and analogues. MIC values are given against MDR-TB, XDR-TB, MRSA, and/or VRE strains. IC<sub>50</sub> values are given for MraY inhibition.



inhibition of phospho-MurNAc-pentapeptide translocase (MraY translocase I). MraY is essential for bacterial cell growth, thus making it a target of interest for the treatment of tuberculosis (TB),<sup>[100]</sup> and even vancomycin- and methicillin-resistant *Staphylococcus aureus* (VRSA, MRSA).<sup>[101]</sup>

Although a very promising antibacterial treatment, CPZ-B has remained synthetically challenging, especially in regard to attaching the unstable fatty acid side chain. This fact, combined with an onerous HPLC separation and a poor water-solubility profile, has prevented it from becoming a therapeutically viable candidate.<sup>[102]</sup> Only recently has a total synthesis been reported for the less-active caprazamycin A<sup>[103]</sup> by Takemoto and co-workers,<sup>[104]</sup> while Watanabe and co-workers have reported a partial synthesis of the western portion of CPZ-B.<sup>[105]</sup> Thus, the primary focus over the years has been on analogues of CPZ, developed primarily by Ichikawa, Matsuda, and co-workers.<sup>[106]</sup> During their studies, the potency of the diketopiperazine and acyclic analogues revealed that the diazepanone moiety was not essential, but contributory to antibacterial activity.<sup>[106b,107]</sup> Replacing the diazepanone and aminoribose units with an oxazolidine moiety restricted the flexibility sufficiently to lead to an increase in potency of MIC = 2–16  $\mu\text{g mL}^{-1}$  against MRSA and VRE (vancomycin-resistant enterococci) strains (oxazolidine analogue; Figure 10).<sup>[107]</sup> The most recent studies include CPZ analogues such as CPZEN-45 with particularly high potency against MDR-TB strains.<sup>[102]</sup> As the oxazolidine analogues were revealed to be weak MraY inhibitors ( $\text{IC}_{50}$  = 920–1200  $\mu\text{M}$ ), Ichikawa et al. recently reported the carbacaprazamycin analogues, with MIC values of 4–16  $\mu\text{g mL}^{-1}$  and MraY inhibition with  $\text{IC}_{50}$  values of 2.6–6.9  $\text{nmol L}^{-1}$ .<sup>[108]</sup> However, the morphological changes observed in *S. aureus* indicated a mode of action that may be completely different from existing peptidoglycan inhibitors.

The most potent oxazolidine analogue **65** can be accessed in just 12 steps, starting with an oxidation, two-carbon elongation, and an aminohydroxylation<sup>[109]</sup> of uridine **62** to afford amino alcohol **63** (Scheme 8).<sup>[107]</sup> Protecting group manipulations and installation of the oxaziridine ring by condensation with azidoacetaldehyde, followed by acylation afforded *N*-palmitoyloxazolidine **64**.<sup>[107]</sup> Finally, conversion of the methyl ester into the *tert*-butyl ester by using an isourea derivative followed by reduction of the azide to the amine afforded oxazolidine analogue **65**. The overall reduction in the complexity of this analogue compared to the parent compound is striking, all the while retaining its potent antibiotic activity against multidrug-resistant strains.

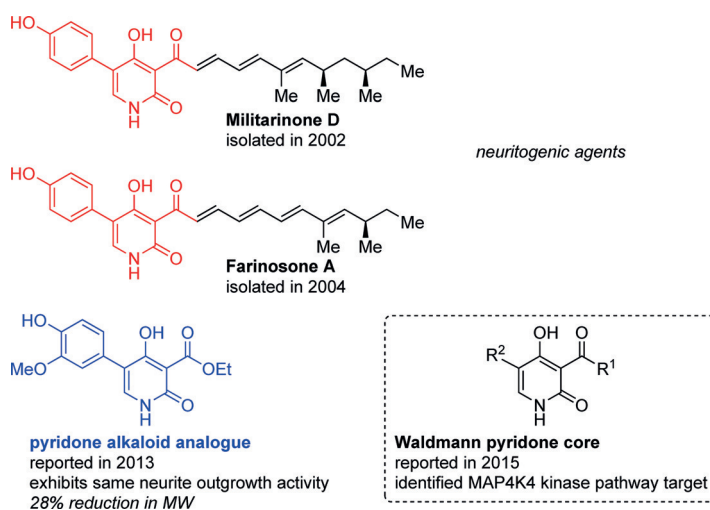


**Scheme 8.** Synthesis of the oxazolidine analogue (**65**) by Ichikawa, Matsuda, and co-workers. Cbz = benzyloxycarbonyl,  $(\text{DHQD})_2\text{AQN}$  = 1,4-bis(dihydroquinidyl)-anthraquinone, IBX = 2-iodoxybenzoic acid.

### 2.3. Neuritogenic Activity

#### 2.3.1. From Militarinone D/Farinosone A to Pyridone Analogues

A plethora of biologically active pyridone alkaloids have been isolated from entomopathogenic *Deuteromycetes* fungi. Militarinone D was isolated in 2002<sup>[110]</sup> and later identified together with several related molecules in 2003<sup>[111]</sup> by Hamburger and co-workers through bioassay-guided fractionation of a mycelial extract of *Paecilomyces militaris* (RCEF 0095), which displayed pronounced neuritogenic activity in PC-12 cells (Figure 11). Isolated from a mycelial extract of a related

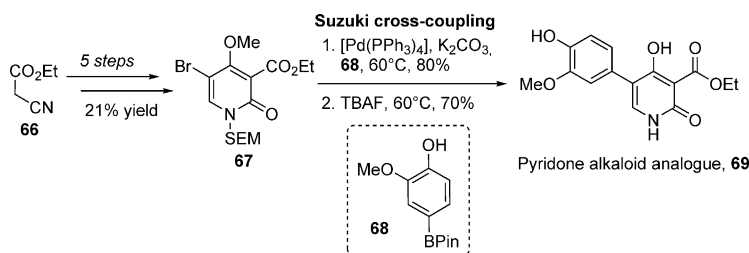


**Figure 11.** Structural comparison of militarinone D and farinosone A with pyridone alkaloid analogues.

fungus, farinosones A and B structurally resemble the militarinones, retain similar neurite outgrowth activity, and exhibit no appreciable cytotoxicity.<sup>[112]</sup> A number of other pyridone alkaloids have been synthesized and evaluated in terms of their biological activity.<sup>[111,113]</sup>

In 2011, Gademann and co-workers developed a unified approach toward several related pyridone alkaloid natural

products.<sup>[114]</sup> They found that these natural products, their enantiomers, and some additional isomers with a *Z*-configured double bond in the side chain demonstrated neuritogenic activity in a standardized PC-12 assay, thereby revealing that neither the length of the side chain nor the absolute configuration were essential to the pharmacophore of this molecule family. In 2013, the Gademann group followed up with further SAR studies and found truncated analogue **69** to have a comparable neurite outgrowth capacity as farinosone A at 20  $\mu\text{M}$ .<sup>[115]</sup> Easily synthesized by utilizing the same general route as for the pyridone natural products, pyridone analogue **69** was accessed in just seven steps from ethyl cyanoacetate (**66**) and 12% overall yield by utilizing a Suzuki–Miyaura coupling between the brominated pyridone **67** and pinacol borane **68** (Scheme 9). The neuritogenic effects of pyridone analogue **69** could even be observed down to concentrations of 1  $\mu\text{M}$ . Further investigations mirrored prior biological studies<sup>[116]</sup> and revealed that this potent analogue, as well as militarinone D, influence the MAP kinase pathway, as the neuritogenic effects could be blocked by co-incubation with the ERK1/2 inhibitor PD98059 (at 5  $\mu\text{M}$ ). Recently, Waldmann and co-workers reported a militarinone-inspired collection of pyridones, and identified the stress pathway kinase MAP4K4 as the target (Figure 11).<sup>[117]</sup> Both

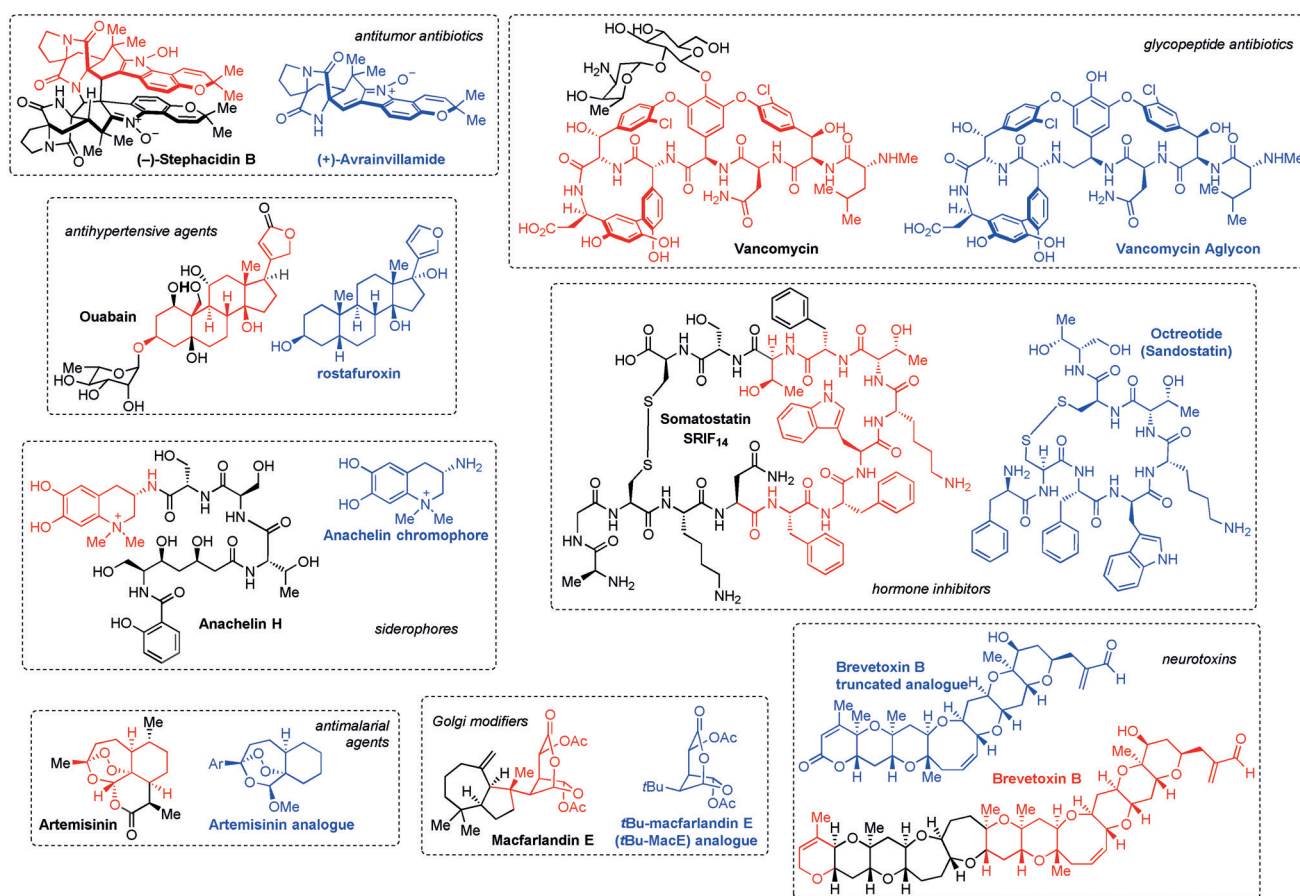


**Scheme 9.** Synthesis of pyridone analogue **69** by Gademann and co-workers. Pin = Pinacol, SEM = trimethylsilylethoxymethyl.

examples show that by reducing the number of carbon atoms by nearly a half, much more simplified, but still potent natural product fragments could be successfully produced.

#### 2.4. Miscellaneous Biological Activity

In addition to those presented within this Review, there are many other notable examples of biologically active, natural product derived fragments (Figure 12). Myers and Herzon demonstrated that avrainvillamide had the same nanomolar antiproliferative activity ( $\text{IC}_{50} = 50\text{--}100\text{ nmol L}^{-1}$ ) against several cancer cell lines as its structural dimer, stephacidin B.<sup>[118]</sup> These investigations, including SAR



**Figure 12.** Structural comparison of additional natural product derived, biologically active fragments and their parent compounds.

studies of several analogues, established the absolute stereochemical configuration, found that this reversible dimerization occurs through the 3-alkylidene-3*H*-indole 1-oxide functional group, attributed the activity of stephacidin B to arise from dissociation to avrainvillamide (as hypothesized), and identified the nucleolar phosphoprotein nucleophosmin as a biological target. The elegant studies of the well-known antibiotic vancomycin and its aglycon by Crowley and Boger found the simplified structure to have more potent activity against a vancomycin-resistant strain than the parent compound ( $\mu\text{g mL}^{-1}$  versus  $\text{mg mL}^{-1}$  activity).<sup>[119]</sup> These investigations reaffirmed the dimeric binding model<sup>[120]</sup> and addressed the growing vancomycin-resistance issue. Another example is the truncation of the 14-mer cyclic peptide hormone somatostatin (SRIF<sub>14</sub>) to the 8-mer, octreotide.<sup>[6c, 121]</sup> Now marketed by Novartis as the acetate salt, this drug is 70 times more potent than its parent peptide in vivo, and is used in the treatment of acromegaly as well as symptoms associated with metastatic carcinoid tumors or vasoactive intestinal peptide-secreting adenomas. This example showed that it is possible to mimic the activity of natural hormones with a truncated, synthetic analogue, and resulted in a clinically successful drug that improves thousands of lives annually. Another clinically relevant example is that of rostauroxin, which is a simplified version of the toxic cardiac glycoside ouabain.<sup>[122]</sup> This antihypertensive agent is currently in phase IIb clinical trials and has been proven to be especially effective for those with genetic abnormalities of adducin and EO (endogenous ouabain) blood pressure regulation. Gademann and co-workers demonstrated that the known siderophore aachelin H could be truncated to its chromophore and reapplied as an anchor to create antifouling and antimicrobial surface coatings for potential use in biocompatible medical devices.<sup>[123]</sup> In 1994, Nicolaou et al. designed and synthesized truncated brevetoxin analogue [AFGHJK], which was able to bind to the voltage-gated sodium channel and exhibit the same electrophysiological activities as the parent, “red-tide” ladder polyether toxin with an additional four rings.<sup>[119b, 124]</sup> More recently, Overman and co-workers reported the *t*Bu-MacE analogue to have the same Golgi-modifying properties as the parent spongian diterpene macfarlandin E, but with even less cytotoxicity.<sup>[125]</sup>

Our last example is from the studies by Posner and O'Neill on analogues of the sesquiterpene trioxane artemisinin. They have reported many simplified aryl derivatives with comparable in vitro antimalarial activities in the nanomolar range, which also function by the iron-induced triggering mechanism and were furthermore effective in rodent models.<sup>[126]</sup>

### 3. Additional Sources of Natural Product Fragment Lead Structures

This Review has focused on the identification of suitable fragments through chemical synthesis. Looking toward the future, however, recent advancements in structural biology, computational chemistry, and genome mining show the

potential to identify suitable natural product fragments that might be further refined by chemical synthesis.

A different approach to access novel, natural product fragments based on genome data is fueled by advancements in fields such as molecular biology, bioinformatics, genetic mapping, and structural biology. Within the realm of nature, DNA-encoding fragments of compounds are exchanged between organisms and incorporated in new structures, which can sometimes be found in different kingdoms.<sup>[127]</sup> The natural product fragments are exchanged within organisms by horizontal gene or fragment gene transfer. A representative example is found in the pederin, theopederin A, and onnamide A family of natural products, which share common fragments.<sup>[128]</sup> The revolution in genome mining and advances in metabolomics will continue to unearth even more natural product structures, at an ever-increasing speed.

Anticancer antibody–drug conjugates (ADCs) are currently the biopharmaceutical standard for innovative cancer treatments, with over 30 entities in clinical development.<sup>[129]</sup> As the name suggests, an antibody is linked to a cytotoxic, natural product derived “payload” to maximize targeting and efficacy of the anticancer agent toward the diseased tissue. Lead optimization in this field involves the development of chemical conjugation methods to couple the natural product to the antibody, the design of the linker, and the type of antibody used.<sup>[130]</sup> Gemtuzumab ozogamicin (Mylotarg) was the first clinically approved ADC. It was launched in 2000 in the USA for the treatment of refractory acute myeloid leukemia (AML), specifically for cells bearing the CD33 antigen.<sup>[129a, 131]</sup> It contains an *N*-acetyl- $\gamma$ -calicheamicin dimethyl hydrazide derivative as its active agent, linked by a pH-labile hydrazone fragment to a recombinant, humanized IgG4  $\kappa$  antibody, developed by Wyeth and UCB Pharma. This success has led to the evaluation of other cytotoxins for this type of cancer chemotherapy, including taxanes, ansamycins, and duocarmycins.<sup>[130]</sup>

More recently, natural product derived fragments are leading the next generation of ADCs for clinical development.<sup>[129a]</sup> There are currently 33 ADCs with natural product derived warheads, of which 10 are in phase II trials and 23 in phase I trials. They are derivatives of calicheamicin  $\gamma$ 1, dolastatin 10 (monomethylaurisatin E and F, MMAE and MMAF), maytansine (DM1 and DM4), doxorubicin, SN-38 (camptothecin, irinotecan derivative), and anthramycin (pyrrolobenzodiazepine, or PBD, dimer; SGD-1882). It is an exciting time in this relatively new field, as the technologies continue to develop for the efficient production of these natural product derived ADCs. Successful lead optimization will likely launch this class of anticancer chemotherapies into predominant clinical use, and eventually application toward treatments of other diseases.

Although fragment-based drug design (FBDD) or screening libraries from diversity-oriented synthesis (DOS) seemed to be the contending methods to approach drug development in recent years, in the past 2–3 years we have seen a novel tactic that combines these two methods and involves computational analysis and data mining to create or repurpose natural product derived fragments (NPDFs).<sup>[132]</sup> This allows for an elegant and systematic identification of biological



targets for “orphan” natural products and/or NPDPs, which can then be validated through chemical synthesis and subsequent biological assessment. In the pioneering report by Waldmann and co-workers in 2013, they disclosed an innovative algorithm in which 2000 clusters of natural NPDPs were generated from 180 000 natural products, rich in  $sp^3$ -hybridized centers.<sup>[133]</sup> The overall concept was to identify unexplored ligand classes and areas of chemical space for established drug targets, especially those that have been difficult to address. This approach combined the advantages of the chemical space offered by fragments with the rich chemistry and geometry that natural products bring, meanwhile leaving a possibility for optimization for biological recognition and modulation.<sup>[132]</sup> As a proof-of-concept, they screened and identified novel scaffolds as inhibitors of p38 $\alpha$  MAP kinase and tyrosine/dual-specificity phosphatases by using protein X-ray crystallography. The term “biology-oriented synthesis” (BIOS) has been coined by Waldmann and co-workers for this method.<sup>[134]</sup>

In another example from 2014, Schneider and co-workers developed a ligand-based model to predict biological targets for NPDPs that does not require a three-dimensional target model, but instead relies on the hypothetical polypharmacological character of natural products.<sup>[135]</sup> To validate their method, they analyzed the antitumor agent archazolid A, and identified eight new biochemical targets for this molecule, all of which have been associated with antitumor effects. Additional work from the Schneider group has focused on repurposing of de novo created compounds from computational models to predict bioactivity,<sup>[136]</sup> or on chemography by creating a generative topographic map (GTM) to visualize chemical space populated by natural products and synthetic drugs.<sup>[137]</sup>

In 2015, Quinn and co-workers demonstrated that their self-generated NPDP library captured more than half of the small pharmacophore triplet diversity than other natural product datasets,<sup>[138]</sup> and Lanz and Riedl used NPDPs as seeds for the de novo discovery of lead structures for therapeutically relevant targets.<sup>[139]</sup>

Overall, these examples delineate a future for this field by proving that the combination of bioinformatic/cheminformatic tools with organic synthesis will enable scientists to expand the boundaries of biologically relevant chemical space.

#### 4. Summary and Outlook

In this Review, we aimed to highlight how a desired property of a natural product, such as its powerful biological activity, can be retained by a smaller, truncated compound (“reduce to the max”)—a refined fragment of the parent natural product. This approach addresses some of the shortcomings of the use of natural products in drug discovery. The chemical tractability can be greatly improved by structurally more simple analogues, with fewer stereogenic centers and often an overall drastically shorter synthetic route. In fact, some of the resulting fragments resemble classical medicinal chemistry targets with regard to size and complexity. In

addition, although considerable advances have been made in synthetic methods over the last decades, chemical synthesis alone cannot likely keep up with the de novo creation of interesting natural product like scaffolds. As a result, a number of potential avenues have opened up the future of this field, including the use of natural product derived fragments in antibody–drug conjugates, stereochemically complex modules for fragment-based drug discovery, or scaffold repurposing, as the next generation of natural product-inspired therapies.

These, among other advancements in such fields as structural biology, genetic mapping, protein crystallography, computational chemistry, nanotechnology, and genome mining show how natural product fragments can be used to address some of the foreseeable challenges of innovative drug development. All these combined approaches clearly present a promising future role for natural products in drug development. Innovation in these trajectories is key to the future of natural product diversity having an impact on and expanding the breadth of the drug discovery field.

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- [1] a) E. S. Istvan, J. Deisenhofer, *Science* **2001**, *292*, 1160–1164; b) K. S. Jain, M. K. Kathiravan, R. S. Somani, C. J. Shishoo, *Bioorg. Med. Chem.* **2007**, *15*, 4674–4699.
- [2] a) J. Szychowski, J. F. Truchon, Y. L. Bennani, *J. Med. Chem.* **2014**, *57*, 9292–9308; b) D. J. Newman, G. M. Cragg, *J. Nat. Prod.* **2012**, *75*, 311–335.
- [3] M. Feher, J. M. Schmidt, *J. Chem. Inf. Comput. Sci.* **2003**, *43*, 218–227.
- [4] For additional reviews, see a) A. M. Szpilman, E. M. Carreira, *Angew. Chem. Int. Ed.* **2010**, *49*, 9592–9628; *Angew. Chem.* **2010**, *122*, 9786–9823; b) P. A. Wender, A. C. Donnelly, B. A. Loy, K. E. Near, D. Staveness in *Natural Products in Medicinal Chemistry, 1st ed.* (Ed.: S. Hanessian), Wiley-VCH, Weinheim, **2014**, pp. 475–543.
- [5] R. M. Wilson, S. J. Danishefsky, *J. Org. Chem.* **2006**, *71*, 8329–8351.
- [6] a) P. A. Wender, V. A. Verma, T. J. Paxton, T. H. Pillow, *Acc. Chem. Res.* **2008**, *41*, 40–49; b) J. Y. Wach, K. Gademann, *Synlett* **2012**, 163–170; c) P. A. Wender, *Tetrahedron* **2013**, *69*, 7529–7550; d) P. A. Wender, R. V. Quiroz, M. C. Stevens, *Acc. Chem. Res.* **2015**, *48*, 752–760.
- [7] a) D. A. Erlanson, R. S. McDowell, T. O'Brien, *J. Med. Chem.* **2004**, *47*, 3463–3482; b) C. W. Murray, D. C. Rees, *Nat. Chem.* **2009**, *1*, 187–192; c) D. E. Scott, A. G. Coyne, S. A. Hudson, C. Abell, *Biochemistry* **2012**, *51*, 4990–5003; d) S. R. Bathula, S. M. Akondi, P. S. Mainkar, S. Chandrasekhar, *Org. Biomol. Chem.* **2015**, *13*, 6432–6448.

- [8] For extensive reviews, see a) K. L. Jackson, J. A. Henderson, A. J. Phillips, *Chem. Rev.* **2009**, *109*, 3044–3079; b) M. J. Yu, Y. Kishi, B. A. Littlefield in *Anticancer Agents from Natural Products* (Eds.: G. M. Cragg, D. G. I. Kingston, D. J. Newman), CRC, Taylor & Francis, Boca Raton, **2012**, pp. 317–345.
- [9] a) D. Uemura, K. Takahashi, T. Yamamoto, C. Katayama, J. Tanaka, Y. Okumura, Y. Hirata, *J. Am. Chem. Soc.* **1985**, *107*, 4796–4798; b) Y. Hirata, D. Uemura, *Pure Appl. Chem.* **1986**, *58*, 701–710.
- [10] G. R. Pettit, C. L. Herald, M. R. Boyd, J. E. Leet, C. Dufresne, D. L. Doubek, J. M. Schmidt, R. L. Cerny, J. N. Hooper, K. C. Rutzler, *J. Med. Chem.* **1991**, *34*, 3339–3340.
- [11] T/C = size of treated tumor/size of control tumor.
- [12] R. L. Bai, K. D. Paull, C. L. Herald, L. Malspeis, G. R. Pettit, E. Hamel, *J. Biol. Chem.* **1991**, *266*, 15882–15889.
- [13] a) M. H. Munro, J. W. Blunt, E. J. Dumdei, S. J. Hickford, R. E. Lill, S. Li, C. N. Battershill, A. R. Duckworth, *J. Biotechnol.* **1999**, *70*, 15–25; b) J. B. Hart, R. E. Lill, S. J. H. Hickford, J. W. Blunt, M. H. G. Munro in *Drugs from the Sea* (Ed.: N. Fusetani), Karger, Basel, **2000**, pp. 134–153; c) Ref. [8b].
- [14] a) J. S. Brimacombe, A. M. Mofti, L. C. N. Tucker, *J. Chem. Soc. C* **1971**, 2911–2915; b) M. Y. H. Wong, G. R. Gray, *J. Am. Chem. Soc.* **1978**, *100*, 3548–3553; c) J. A. J. M. Vekemans, J. Boerekamp, E. F. Godefroi, G. J. F. Chittenden, *Recl. Trav. Chim. Pays-Bas* **1985**, *104*, 266–272; d) T. D. Aicher, Y. Kishi, *Tetrahedron Lett.* **1987**, *28*, 3463–3466; e) T. D. Aicher, K. R. Buszek, F. G. Fang, C. J. Forsyth, S. H. Jung, Y. Kishi, M. C. Matelich, P. M. Scola, D. M. Spero, S. K. Yoon, *J. Am. Chem. Soc.* **1992**, *114*, 3162–3164; f) T. D. Aicher, K. R. Buszek, F. G. Fang, C. J. Forsyth, S. H. Jung, Y. Kishi, P. M. Scola, *Tetrahedron Lett.* **1992**, *33*, 1549–1552; g) K. R. Buszek, F. G. Fang, C. J. Forsyth, S. H. Jung, Y. Kishi, P. M. Scola, S. K. Yoon, *Tetrahedron Lett.* **1992**, *33*, 1553–1556.
- [15] a) K. Takai, K. Kimura, T. Kuroda, T. Hiyama, H. Nozaki, *Tetrahedron Lett.* **1983**, *24*, 5281–5284; b) N. A. Hughes, *Carbohydr. Res.* **1968**, *7*, 474–479; c) B. B. Lohray, T. H. Kalantar, B. M. Kim, C. Y. Park, T. Shibata, J. S. M. Wai, K. B. Sharpless, *Tetrahedron Lett.* **1989**, *30*, 2041–2044; d) K. Tomooka, T. Okinaga, K. Suzuki, G. Tsuchihashi, *Tetrahedron Lett.* **1989**, *30*, 1563–1566; e) E. W. Colvin, B. J. Hamill, *J. Chem. Soc. Chem. Commun.* **1973**, 151–152; f) E. W. Colvin, B. J. Hamill, *J. Chem. Soc. Perkin Trans. 1* **1977**, 869–874; g) J. C. Gilbert, U. Weerasooriya, *J. Org. Chem.* **1979**, *44*, 4997–4998; h) J. Inanaga, K. Hirata, H. Saeki, T. Katsuki, M. Yamaguchi, *Bull. Chem. Soc. Jpn.* **1979**, *52*, 1989–1993.
- [16] a) J. J. W. Duan, Y. Kishi, *Tetrahedron Lett.* **1993**, *34*, 7541–7544; b) D. P. Stamos, Y. Kishi, *Tetrahedron Lett.* **1996**, *37*, 8643–8646; c) S. B. Liu, J. T. Kim, C. G. Dong, Y. Kishi, *Org. Lett.* **2009**, *11*, 4520–4523; d) D. P. Stamos, S. S. Chen, Y. Kishi, *J. Org. Chem.* **1997**, *62*, 7552–7553; e) K. Namba, H. S. Jun, Y. Kishi, *J. Am. Chem. Soc.* **2004**, *126*, 7770–7771.
- [17] a) Y. Wang, G. J. Habgood, W. J. Christ, Y. Kishi, B. A. Littlefield, M. J. Yu, *Bioorg. Med. Chem. Lett.* **2000**, *10*, 1029–1032; b) B. M. Seletsky, Y. Wang, L. D. Hawkins, M. H. Palme, G. J. Habgood, L. V. DiPietro, M. J. Towle, K. A. Salvato, B. F. Wels, K. K. Aalfs, Y. Kishi, B. A. Littlefield, M. J. Yu, *Bioorg. Med. Chem. Lett.* **2004**, *14*, 5547–5550.
- [18] a) M. J. Towle, K. A. Salvato, J. Budrow, B. F. Wels, G. Kuznetsov, K. K. Aalfs, S. Welsh, W. Zheng, B. M. Seletsky, M. H. Palme, G. J. Habgood, L. A. Singer, L. V. DiPietro, Y. Wang, J. J. Chen, D. A. Quincy, A. Davis, K. Yoshimatsu, Y. Kishi, M. J. Yu, B. A. Littlefield, *Cancer Res.* **2001**, *61*, 1013–1021; b) W. Zheng, B. M. Seletsky, M. H. Palme, P. J. Lydon, L. A. Singer, C. E. Chase, C. A. Lemelin, Y. Shen, H. Davis, L. Tremblay, M. J. Towle, K. A. Salvato, B. F. Wels, K. K. Aalfs, Y. Kishi, B. A. Littlefield, M. J. Yu, *Bioorg. Med. Chem. Lett.* **2004**, *14*, 5551–5554.
- [19] a) D. S. Kim, C. G. Dong, J. T. Kim, H. B. Guo, J. Huang, P. S. Tiseni, Y. Kishi, *J. Am. Chem. Soc.* **2009**, *131*, 15636–15641; b) Y. R. Yang, D. S. Kim, Y. Kishi, *Org. Lett.* **2009**, *11*, 4516–4519; c) M. D. Shan, Y. Kishi, *Org. Lett.* **2012**, *14*, 660–663.
- [20] B. C. Austad, F. Benayoud, T. L. Calkins, S. Campagna, C. E. Chase, H. W. Choi, W. Christ, R. Costanzo, J. Cutter, A. Endo, F. G. Fang, Y. B. Hu, B. M. Lewis, M. D. Lewis, S. McKenna, T. A. Noland, J. D. Orr, M. Pesant, M. J. Schnaderbeck, G. D. Wilkie, T. Abe, N. Asai, Y. Asai, A. Kayano, Y. Kimoto, Y. Komatsu, M. Kubota, H. Kuroda, M. Mizuno, T. Nakamura, T. Omae, N. Ozeki, T. Suzuki, T. Takigawa, T. Watanabe, K. Yoshizawa, *Synlett* **2013**, *24*, 327–332.
- [21] M. Tokunaga, J. F. Larrow, F. Kakiuchi, E. N. Jacobsen, *Science* **1997**, *277*, 936–938.
- [22] M. Kurosu, M. H. Lin, Y. Kishi, *J. Am. Chem. Soc.* **2004**, *126*, 12248–12249.
- [23] See Ref. 18b.
- [24] Y. Kaburagi, Y. Kishi, *Tetrahedron Lett.* **2007**, *48*, 8967–8971.
- [25] Y. Kaburagi, Y. Kishi, *Org. Lett.* **2007**, *9*, 723–726.
- [26] C. G. Dong, J. A. Henderson, Y. Kaburagi, T. Sasaki, D. S. Kim, J. T. Kim, D. Urabe, H. Guo, Y. Kishi, *J. Am. Chem. Soc.* **2009**, *131*, 15642–15646.
- [27] a) FDA approves new treatment option for late-stage breast cancer, press release, U.S. Food and Drug Administration, **2010**; available from: <http://www.fda.gov/NewsEvents/Newsroom/PressAnnouncements/ucm233863.htm>, last accessed: October 12, **2015**; b) *E7389 Versus Treatment of Physician's Choice in Patients With Locally Recurrent or Metastatic Breast Cancer*, U.S. National Institutes of Health, **2006**; available from: <https://clinicaltrials.gov/show/NCT00388726> identifier: NCT00388726, last accessed: January 27, **2015**.
- [28] Eisai Receives European Commission Approval of Indication Expansion for Anticancer Agent Halaven for Advanced Breast cancer After Only One Prior Chemotherapy, press release, Eisai, **2014**; available from: <http://www.eisai.com/news/news201437.html>, last accessed: October 12, **2015**.
- [29] a) K. Nakae, Y. Yoshimoto, T. Sawa, Y. Homma, M. Hamada, T. Takeuchi, M. Imoto, *J. Antibiot.* **2000**, *53*, 1130–1136; b) K. Nakae, Y. Yoshimoto, M. Ueda, T. Sawa, Y. Takahashi, H. Naganawa, T. Takeuchi, M. Imoto, *J. Antibiot.* **2000**, *53*, 1228–1230; c) Y. Takemoto, K. Nakae, M. Kawatani, Y. Takahashi, H. Naganawa, M. Imoto, *J. Antibiot.* **2001**, *54*, 1104–1107.
- [30] H. Nakamura, Y. Takahashi, H. Naganawa, K. Nakae, M. Imoto, M. Shiro, K. Matsumura, H. Watanabe, T. Kitahara, *J. Antibiot.* **2002**, *55*, 442–444.
- [31] a) C. Gaul, S. J. Danishefsky, *Tetrahedron Lett.* **2002**, *43*, 9039–9042; b) C. Gaul, J. T. Njardarson, S. J. Danishefsky, *J. Am. Chem. Soc.* **2003**, *125*, 6042–6043; c) C. Gaul, J. T. Njardarson, D. Shan, D. C. Dorn, K. D. Wu, W. P. Tong, X. Y. Huang, M. A. S. Moore, S. J. Danishefsky, *J. Am. Chem. Soc.* **2004**, *126*, 11326–11337; d) for additional synthetic studies towards migrastatin, see e) S. Reymond, J. Cossy, *Eur. J. Org. Chem.* **2006**, 4800–4804; f) S. Reymond, J. Cossy, *Tetrahedron* **2007**, *63*, 5918–5929; g) G. Anquetin, G. Horgan, S. Rawe, D. Murray, A. Madden, P. MacMathuna, P. Doran, P. V. Murphy, *Eur. J. Org. Chem.* **2008**, 1953–1958; h) J. S. Yadav, P. N. Lakshmi, *Synlett* **2010**, 1033–1036; i) L. C. Dias, F. G. Finelli, L. S. Conejero, R. Krogh, A. D. Andricopulo, *Eur. J. Org. Chem.* **2009**, 6748–6759; j) N. R. Gade, J. Iqbal, *Tetrahedron Lett.* **2013**, *54*, 4225–4227; k) N. R. Gade, J. Iqbal, *Eur. J. Org. Chem.* **2014**, 6558–6564.
- [32] a) J. T. Njardarson, C. Gaul, D. Shan, X. Y. Huang, S. J. Danishefsky, *J. Am. Chem. Soc.* **2004**, *126*, 1038–1040; b) T. Oskarsson, P. Nagorny, I. J. Krauss, L. Perez, M. Mandal, G. L. Yang, O. Ouerfelli, D. H. Xiao, M. A. S. Moore, J. Massague, S. J. Danishefsky, *J. Am. Chem. Soc.* **2010**, *132*, 3224–3228.

- [33] N. Lecomte, J. T. Njardarson, P. Nagorny, G. L. Yang, R. Downey, O. Ouerfelli, M. A. S. Moore, S. J. Danishefsky, *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 15074–15078.
- [34] a) D. D. Shan, L. Chen, J. T. Njardarson, C. Gaul, X. J. Ma, S. J. Danishefsky, X. Y. Huang, *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 3772–3776; b) additional studies also found that the migrastatin macroketone analogue also significantly increase the immobile fraction E-cadherin in vivo by FRAP (fluorescence recovery after photobleaching) in 2014 by Murphy and co-workers; see D. Lo Re, Y. Zhou, M. Nobis, K. I. Anderson, P. V. Murphy, *ChemBioChem* **2014**, *15*, 1459–1464.
- [35] L. Chen, S. Yang, J. Jakoncic, J. J. Zhang, X. Y. Huang, *Nature* **2010**, *464*, 1062–1066.
- [36] P. Nagorny, I. Krauss, J. T. Njardarson, L. Perez, C. Gaul, G. L. Yang, O. Ouerfelli, S. J. Danishefsky, *Tetrahedron Lett.* **2010**, *51*, 3873–3875.
- [37] K. Majchrzak, D. Lo Re, M. Gajewska, M. Bulkowska, A. Homa, K. Pawlowski, T. Motyl, P. V. Murphy, M. Krol, *PLoS One* **2013**, *8*, e76789.
- [38] Y. Hayakawa, K. Sohda, K. Shinya, T. Hidaka, H. Seto, *J. Antibiot.* **1995**, *48*, 954–961.
- [39] S. Bonazzi, S. Guttinger, I. Zemp, U. Kutay, K. Gademann, *Angew. Chem. Int. Ed.* **2007**, *46*, 8707–8710; *Angew. Chem.* **2007**, *119*, 8862–8865.
- [40] a) N. Kudo, B. Wolff, T. Sekimoto, E. P. Schreiner, Y. Yoneda, M. Yanagida, S. Horinouchi, M. Yoshida, *Exp. Cell Res.* **1998**, *242*, 540–547; b) N. Kudo, N. Matsumori, H. Taoka, D. Fujiwara, E. P. Schreiner, B. Wolff, M. Yoshida, S. Horinouchi, *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 9112–9117; c) X. H. Dong, A. Biswas, Y. M. Chook, *Nat. Struct. Mol. Biol.* **2009**, *16*, 558–560; d) X. H. Dong, A. Biswas, K. E. Suel, L. K. Jackson, R. Martinez, H. M. Gu, Y. M. Chook, *Nature* **2009**, *458*, 1136–1141; e) T. Monecke, T. Guttler, P. Neumann, A. Dickmanns, D. Gorlich, R. Ficner, *Science* **2009**, *324*, 1087–1091.
- [41] Q. X. Sun, Y. P. Carrasco, Y. C. Hu, X. F. Guo, H. Mirzaei, J. MacMillan, Y. M. Chook, *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 1303–1308.
- [42] E. S. Newlands, G. J. S. Rustin, M. H. Brampton, *Br. J. Cancer* **1996**, *74*, 648–649.
- [43] S. Bonazzi, O. Eidam, S. Guttinger, J. Y. Wach, I. Zemp, U. Kutay, K. Gademann, *J. Am. Chem. Soc.* **2010**, *132*, 1432–1442.
- [44] M. J. Plater, S. Aiken, G. Bourhill, *Tetrahedron* **2002**, *58*, 2415–2422.
- [45] K. Komatsu, K. Tanino, M. Miyashita, *Angew. Chem. Int. Ed.* **2004**, *43*, 4341–4345; *Angew. Chem.* **2004**, *116*, 4441–4445.
- [46] T. Hintermann, D. Seebach, *Helv. Chim. Acta* **1998**, *81*, 2093–2126.
- [47] a) J. A. Marshall, M. P. Bourbeau, *J. Org. Chem.* **2002**, *67*, 2751–2754; b) J. A. Marshall, A. M. Mikowski, M. P. Bourbeau, G. M. Schaaf, F. Valeriote, *Bioorg. Med. Chem. Lett.* **2006**, *16*, 320–323.
- [48] Adapted from Ref. 43 with permission.
- [49] C. Kalyanaraman, K. Bernacki, M. P. Jacobson, *Biochemistry* **2005**, *44*, 2059–2071.
- [50] K. Gademann, *Curr. Drug Targets* **2011**, *12*, 1574–1580.
- [51] M. Ichimura, T. Ogawa, K. Takahashi, E. Kobayashi, I. Kawamoto, T. Yasuzawa, I. Takahashi, H. Nakano, *J. Antibiot.* **1990**, *43*, 1037–1038.
- [52] a) D. L. Boger, D. S. Johnson, *Proc. Natl. Acad. Sci. USA* **1995**, *92*, 3642–3649; b) K. S. MacMillan, D. L. Boger, *J. Med. Chem.* **2009**, *52*, 5771–5780.
- [53] D. L. Boger, T. Ishizaki, H. Zarrinmayeh, S. A. Munk, P. A. Kitos, O. Suntornwat, *J. Am. Chem. Soc.* **1990**, *112*, 8961–8971.
- [54] a) D. L. Boger, K. Machiya, *J. Am. Chem. Soc.* **1992**, *114*, 10056–10058; b) D. L. Boger, K. Machiya, D. L. Hertzog, P. A. Kitos, D. Holmes, *J. Am. Chem. Soc.* **1993**, *115*, 9025–9036.
- [55] D. L. Boger, R. S. Coleman, *J. Am. Chem. Soc.* **1988**, *110*, 4796–4807.
- [56] E. C. Taylor, D. J. Dumas, *J. Org. Chem.* **1981**, *46*, 1394–1402.
- [57] M. S. Tichenor, K. S. MacMillan, J. S. Stover, S. E. Wolkenberg, M. G. Pavani, L. Zanella, A. N. Zaid, G. Spalluto, T. J. Rayl, I. Hwang, P. G. Baraldi, D. L. Boger, *J. Am. Chem. Soc.* **2007**, *129*, 14092–14099.
- [58] K. S. MacMillan, J. P. Lajiness, C. L. Cara, R. Romagnoli, W. M. Robertson, I. Hwang, P. G. Baraldi, D. L. Boger, *Bioorg. Med. Chem. Lett.* **2009**, *19*, 6962–6965.
- [59] a) M. Konishi, H. Ohkuma, K. Matsumoto, T. Tsuno, H. Kamei, T. Miyaki, T. Oki, H. Kawaguchi, G. D. Vanduyne, J. Clardy, *J. Antibiot.* **1989**, *42*, 1449–1452; b) M. Konishi, H. Ohkuma, T. Tsuno, T. Oki, G. D. Vanduyne, J. Clardy, *J. Am. Chem. Soc.* **1990**, *112*, 3715–3716.
- [60] a) Y. Sugiura, T. Shiraki, M. Konishi, T. Oki, *Proc. Natl. Acad. Sci. USA* **1990**, *87*, 3831–3835; b) P. A. Wender, R. C. Kelly, S. Beckham, B. L. Miller, *Proc. Natl. Acad. Sci. USA* **1991**, *88*, 8835–8839; c) M. F. Semmelhack, J. Gallagher, D. Cohen, *Tetrahedron Lett.* **1990**, *31*, 1521–1522.
- [61] a) J. A. Porco, F. J. Schoenen, T. J. Stout, J. Clardy, S. L. Schreiber, *J. Am. Chem. Soc.* **1990**, *112*, 7410–7411; b) K. C. Nicolaou, C. K. Hwang, A. L. Smith, S. V. Wendeborn, *J. Am. Chem. Soc.* **1990**, *112*, 7416–7418; c) T. Nishikawa, M. Isobe, T. Goto, *Synlett* **1991**, 393–395; d) J. Taunton, J. L. Wood, S. L. Schreiber, *J. Am. Chem. Soc.* **1993**, *115*, 10378–10379; e) T. Y. Yoon, M. D. Shair, S. J. Danishefsky, G. K. Shulte, *J. Org. Chem.* **1994**, *59*, 3752–3754; f) M. D. Shair, T. Y. Yoon, S. J. Danishefsky, *Angew. Chem. Int. Ed. Engl.* **1995**, *34*, 1721–1723; *Angew. Chem.* **1995**, *107*, 1883–1885; g) M. D. Shair, T. Y. Yoon, K. K. Mosny, T. C. Chou, S. J. Danishefsky, *J. Am. Chem. Soc.* **1996**, *118*, 9509–9525.
- [62] a) A. G. Myers, M. E. Fraley, N. J. Tom, S. B. Cohen, D. J. Madar, *Chem. Biol.* **1995**, *2*, 33–43; b) A. G. Myers, M. E. Fraley, N. J. Tom, *J. Am. Chem. Soc.* **1994**, *116*, 11556–11557.
- [63] a) P. A. Wender, C. K. Zercher, *J. Am. Chem. Soc.* **1991**, *113*, 2311–2313; b) P. A. Wender, C. K. Zercher, S. Beckham, E. M. Haubold, *J. Org. Chem.* **1993**, *58*, 5867–5869; c) P. A. Wender, S. Beckham, J. G. O'Leary, *Synthesis* **1994**, 1278–1282; d) P. A. Wender, S. Beckham, D. L. Mohler, *Tetrahedron Lett.* **1995**, *36*, 209–212.
- [64] a) K. C. Nicolaou, W. M. Dai, S. C. Tsay, V. A. Estevez, W. Wrasidlo, *Science* **1992**, *256*, 1172–1178; b) K. C. Nicolaou, A. L. Smith, *Acc. Chem. Res.* **1992**, *25*, 497–503; c) K. C. Nicolaou, W. M. Dai, *J. Am. Chem. Soc.* **1992**, *114*, 8908–8921; d) K. C. Nicolaou, A. L. Smith, E. W. Yue, *Proc. Natl. Acad. Sci. USA* **1993**, *90*, 5881–5888; e) K. C. Nicolaou, W. M. Dai, Y. P. Hong, S. C. Tsay, K. K. Baldrige, J. S. Siegel, *J. Am. Chem. Soc.* **1993**, *115*, 7944–7953.
- [65] a) R. Unno, H. Michishita, H. Inagaki, Y. Suzuki, Y. Baba, T. Jomori, T. Nishikawa, M. Isobe, *Bioorg. Med. Chem.* **1997**, *5*, 883–901; b) R. Unno, H. Michishita, H. Inagaki, Y. Suzuki, Y. Baba, T. Jomori, M. Moku, T. Nishikawa, M. Isobe, *Bioorg. Med. Chem.* **1997**, *5*, 903–919; c) R. Unno, H. Michishita, H. Inagaki, Y. Suzuki, Y. Baba, T. Jomori, T. Nishikawa, M. Isobe, *Bioorg. Med. Chem.* **1997**, *5*, 987–999.
- [66] a) A. G. Myers, S. B. Cohen, N. J. Tom, D. J. Madar, M. E. Fraley, *J. Am. Chem. Soc.* **1995**, *117*, 7574–7575; b) A. G. Myers, N. J. Tom, M. E. Fraley, S. B. Cohen, D. J. Madar, *J. Am. Chem. Soc.* **1997**, *119*, 6072–6094.
- [67] a) M. D. Shair, T. Yoon, T. C. Chou, S. J. Danishefsky, *Angew. Chem. Int. Ed. Engl.* **1994**, *33*, 2477–2479; *Angew. Chem.* **1994**, *106*, 2578–2580; b) S. J. Danishefsky, M. D. Shair, *J. Org. Chem.* **1996**, *61*, 16–44.
- [68] F. Bosse, A. R. Tunoori, M. E. Maier, *Tetrahedron* **1997**, *53*, 9159–9168.



- [69] a) P. Magnus, S. A. Eisenbeis, W. C. Rose, N. Zein, W. Solomon, *J. Am. Chem. Soc.* **1993**, *115*, 12627–12628; b) P. Magnus, S. A. Eisenbeis, R. A. Fairhurst, T. Iliadis, N. A. Magnus, D. Parry, *J. Am. Chem. Soc.* **1997**, *119*, 5591–5605.
- [70] a) M. E. Maier, F. Bosse, A. J. Niestroj, *Eur. J. Org. Chem.* **1999**, 1–13; b) S. C. Sinha, L. S. Li, G. P. Miller, S. Dutta, C. Rader, R. A. Lerner, *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 3095–3099; c) R. Riva, L. Banfi, A. Basso, V. Gandolfo, G. Guanti, *Eur. J. Org. Chem.* **2010**, 2768–2787.
- [71] K. C. Nicolaou, P. Maligres, T. Suzuki, S. V. Wendeborn, W. M. Dai, R. K. Chadha, *J. Am. Chem. Soc.* **1992**, *114*, 8890–8907.
- [72] a) G. R. Pettit, C. L. Herald, D. L. Doubek, D. L. Herald, E. Arnold, J. Clardy, *J. Am. Chem. Soc.* **1982**, *104*, 6846–6848; b) for an extensive review on the byrostatin studies by the Wender group, see P. A. Wender, A. C. Donnelly, B. A. Loy, K. E. Near, D. Staveness in *Natural Products in Medicinal Chemistry, 1st ed.* (Ed.: S. Hanessian), Wiley-VCH, Weinheim, **2014**, pp. 475–543.
- [73] a) M. Hildebrand, L. E. Waggoner, H. B. Liu, S. Sudek, S. Allen, C. Anderson, D. H. Sherman, M. Haygood, *Chem. Biol.* **2004**, *11*, 1543–1552; b) S. Sudek, N. B. Lopanik, L. E. Waggoner, M. Hildebrand, C. Anderson, H. B. Liu, A. Patel, D. H. Sherman, M. G. Haygood, *J. Nat. Prod.* **2007**, *70*, 67–74.
- [74] Z. Szallasi, C. B. Smith, G. R. Pettit, P. M. Blumberg, *J. Biol. Chem.* **1994**, *269*, 2118–2124.
- [75] a) A. W. Elgie, J. M. Sargent, P. Alton, G. J. Peters, P. Noordhuis, C. J. Williamson, C. G. Taylor, *Leuk. Res.* **1998**, *22*, 373–378; b) N. R. Wall, R. M. Mohammad, A. M. Al-Katib, *Leuk. Res.* **1999**, *23*, 881–888; c) H. S. Oz, W. T. Hughes, J. E. Reh, E. K. Thomas, *Microb. Pathog.* **2000**, *29*, 187–190.
- [76] a) J. Kortmansky, G. K. Schwartz, *Cancer Invest.* **2003**, *21*, 924–936; b) S. P. Shaha, J. Tomic, Y. Shi, T. Pham, P. Mero, D. White, L. He, J. L. Baryza, P. A. Wender, J. W. Booth, D. E. Spaner, *Clin. Exp. Immunol.* **2009**, *158*, 186–198; c) for current clinical information, see <http://clinicaltrials.gov> [last accessed: May 2015].
- [77] a) R. Etcheberrigaray, M. Tan, I. Dewachter, C. Kuiperi, I. Van der Auwera, S. Wera, L. X. Qiao, B. Bank, T. J. Nelson, A. P. Kozikowski, F. Van Leuven, D. L. Alkon, *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 11141–11146; b) J. Hongpaisan, D. L. Alkon, *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 19571–19576.
- [78] a) W. M. O. Boto, L. Brown, J. Chrest, W. H. Adler, *Cell Regul.* **1991**, *2*, 95–103; b) B. A. DeChristopher, B. A. Loy, M. D. Marsden, A. J. Schrier, J. A. Zack, P. A. Wender, *Nat. Chem.* **2012**, *4*, 705–710.
- [79] M. Kageyama, T. Tamura, M. H. Nantz, J. C. Roberts, P. Somfai, D. C. Whritenour, S. Masamune, *J. Am. Chem. Soc.* **1990**, *112*, 7407–7408.
- [80] a) D. A. Evans, E. M. Carreira, *Tetrahedron Lett.* **1990**, *31*, 4703–4706; b) D. A. Evans, P. H. Carter, E. M. Carreira, J. A. Prunet, A. B. Charette, M. Lautens, *Angew. Chem. Int. Ed.* **1998**, *37*, 2354–2359; *Angew. Chem.* **1998**, *110*, 2526–2530; c) D. A. Evans, P. H. Carter, E. M. Carreira, A. B. Charette, J. A. Prunet, M. Lautens, *J. Am. Chem. Soc.* **1999**, *121*, 7540–7552.
- [81] K. Ohmori, Y. Ogawa, T. Obitsu, Y. Ishikawa, S. Nishiyama, S. Yamamura, *Angew. Chem. Int. Ed.* **2000**, *39*, 2290–2294; *Angew. Chem.* **2000**, *112*, 2376–2379.
- [82] B. M. Trost, G. B. Dong, *Nature* **2008**, *456*, 485–488.
- [83] G. E. Keck, Y. B. Poudel, T. J. Cummins, A. Rudra, J. A. Covell, *J. Am. Chem. Soc.* **2011**, *133*, 744–747.
- [84] P. A. Wender, A. J. Schrier, *J. Am. Chem. Soc.* **2011**, *133*, 9228–9231.
- [85] Y. Lu, S. K. Woo, M. J. Krische, *J. Am. Chem. Soc.* **2011**, *133*, 13876–13879.
- [86] a) P. A. Wender, K. F. Koehler, N. A. Sharkey, M. L. Dellaquila, P. M. Blumberg, *Proc. Natl. Acad. Sci. USA* **1986**, *83*, 4214–4218; b) P. A. Wender, C. M. Cribbs, K. F. Koehler, N. A. Sharkey, C. L. Herald, Y. Kamano, G. R. Pettit, P. M. Blumberg, *Proc. Natl. Acad. Sci. USA* **1988**, *85*, 7197–7201; c) P. A. Wender, J. DeBrabander, P. G. Harran, J. M. Jimenez, M. F. T. Koehler, B. Lippa, C. M. Park, C. Siedenbiedel, G. R. Pettit, *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 6624–6629; d) K. Kimura, M. Y. Mizutani, N. Tomioka, Y. Endo, K. Shudo, A. Itai, *Chem. Pharm. Bull.* **1999**, *47*, 1134–1137.
- [87] a) P. A. Wender, B. Lippa, *Tetrahedron Lett.* **2000**, *41*, 1007–1011; b) P. A. Wender, M. O. Clarke, J. C. Horan, *Org. Lett.* **2005**, *7*, 1995–1998; c) P. A. Wender, J. C. Horan, *Org. Lett.* **2006**, *8*, 4581–4584; d) P. A. Wender, J. Reuber, *Tetrahedron* **2011**, *67*, 9998–10005; e) P. A. Wender, J. L. Baryza, S. E. Brenner, B. A. DeChristopher, B. A. Loy, A. J. Schrier, V. A. Verma, *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 6721–6726.
- [88] P. A. Wender, J. C. Horan, V. A. Verma, *Org. Lett.* **2006**, *8*, 5299–5302.
- [89] a) P. A. Wender, K. W. Hinkle, *Tetrahedron Lett.* **2000**, *41*, 6725–6729; b) P. A. Wender, J. L. Baryza, *Org. Lett.* **2005**, *7*, 1177–1180.
- [90] P. A. Wender, V. A. Verma, *Org. Lett.* **2008**, *10*, 3331–3334.
- [91] B. M. Trost, H. B. Yang, O. R. Thiel, A. J. Frontier, C. S. Brindle, *J. Am. Chem. Soc.* **2007**, *129*, 2206–2207.
- [92] a) K. J. Hale, M. Frigerio, S. Manaviazar, M. G. Hummersone, I. J. Fillingham, I. G. Barsukov, C. F. Damblon, A. Gescher, G. C. K. Roberts, *Org. Lett.* **2003**, *5*, 499–502; b) G. E. Keck, Y. B. Poudel, D. S. Welch, M. B. Kraft, A. P. Truong, J. C. Stephens, N. Keddi, N. E. Lewin, P. M. Blumberg, *Org. Lett.* **2009**, *11*, 593–596.
- [93] a) P. A. Wender, J. De Brabander, P. G. Harran, J. M. Jimenez, M. F. T. Koehler, B. Lippa, C. M. Park, M. Shiozaki, *J. Am. Chem. Soc.* **1998**, *120*, 4534–4535; b) P. A. Wender, J. L. Baryza, C. E. Bennett, C. Bi, S. E. Brenner, M. O. Clarke, J. C. Horan, C. Kan, E. Lacote, B. Lippa, P. G. Nell, T. M. Turner, *J. Am. Chem. Soc.* **2002**, *124*, 13648–13649; c) P. A. Wender, A. V. W. Mayweg, C. L. VanDeusen, *Org. Lett.* **2003**, *5*, 277–279; d) B. A. DeChristopher, A. C. Fan, D. W. Felsher, P. A. Wender, *Oncotarget* **2012**, *3*, 58–66.
- [94] For a minireview on Prins macrocyclization approaches in polyketide natural product synthesis, see E. A. Crane, K. A. Scheidt, *Angew. Chem. Int. Ed.* **2010**, *49*, 8316–8326; *Angew. Chem.* **2010**, *122*, 8494–8505.
- [95] a) P. A. Wender, B. A. DeChristopher, A. J. Schrier, *J. Am. Chem. Soc.* **2008**, *130*, 6658–6659; b) G. E. Keck, M. B. Kraft, A. P. Truong, W. Li, C. C. Sanchez, N. Keddi, N. E. Lewin, P. M. Blumberg, *J. Am. Chem. Soc.* **2008**, *130*, 6660–6661.
- [96] a) G. E. Keck, Y. B. Poudel, A. Rudra, J. C. Stephens, N. Keddi, N. E. Lewin, M. L. Peach, P. M. Blumberg, *Angew. Chem. Int. Ed.* **2010**, *49*, 4580–4584; *Angew. Chem.* **2010**, *122*, 4684–4688; b) M. B. Kraft, Y. B. Poudel, N. Keddi, N. E. Lewin, M. L. Peach, P. M. Blumberg, G. E. Keck, *J. Am. Chem. Soc.* **2014**, *136*, 13202–13208.
- [97] a) P. A. Wender, Y. Nakagawa, K. E. Near, D. Staveness, *Org. Lett.* **2014**, *16*, 5136–5139; b) P. A. Wender, D. Staveness, *Org. Lett.* **2014**, *16*, 5140–5143.
- [98] M. Igarashi, N. Nakagawa, N. Doi, S. Hattori, H. Naganawa, M. Hamada, *J. Antibiot.* **2003**, *56*, 580–583.
- [99] K. Kimura, N. Miyata, G. Kawanishi, Y. Kamio, K. Izaki, K. Isono, *Agric. Biol. Chem.* **1989**, *53*, 1811–1815.
- [100] C. Dini, *Curr. Top. Med. Chem.* **2005**, *5*, 1221–1236.
- [101] K. Kimura, T. D. H. Bugg, *Nat. Prod. Rep.* **2003**, *20*, 252–273.
- [102] Y. Takahashi, M. Igarashi, T. Miyake, H. Soutome, K. Ishikawa, Y. Komatsuki, Y. Koyama, N. Nakagawa, S. Hattori, K. Inoue, N. Doi, Y. Akamatsu, *J. Antibiot.* **2013**, *66*, 171–178.

- [103] M. Igarashi, Y. Takahashi, T. Shitara, H. Nakamura, H. Naganawa, T. Miyake, Y. Akamatsu, *J. Antibiot.* **2005**, *58*, 327–337.
- [104] H. Nakamura, C. Tsukano, M. Yasui, S. Yokouchi, M. Igarashi, Y. Takemoto, *Angew. Chem. Int. Ed.* **2015**, *54*, 3136–3139; *Angew. Chem.* **2015**, *127*, 3179–3182.
- [105] a) P. Gopinath, T. Watanabe, M. Shibasaki, *J. Org. Chem.* **2012**, *77*, 9260–9267; b) in 2014, another synthesis of (+)-caprazol was published: P. Gopinath, L. Wang, H. Abe, G. Ravi, T. Masuda, T. Watanabe, M. Shibasaki, *Org. Lett.* **2014**, *16*, 3364–3367.
- [106] a) S. Hirano, S. Ichikawa, A. Matsuda, *Angew. Chem. Int. Ed.* **2005**, *44*, 1854–1856; *Angew. Chem.* **2005**, *117*, 1888–1890; b) S. Hirano, S. Ichikawa, A. Matsuda, *J. Org. Chem.* **2008**, *73*, 569–577; c) M. Yamaguchi, A. Matsuda, S. Ichikawa, *Org. Biomol. Chem.* **2015**, *13*, 1187–1197.
- [107] K. Ii, S. Ichikawa, B. Al-Dabbagh, A. Bouhss, A. Matsuda, *J. Med. Chem.* **2010**, *53*, 3793–3813.
- [108] S. Ichikawa, M. Yamaguchi, L. S. Hsuan, A. Matsuda, *ACS Infect. Dis.* **2015**, *1*, 151–156.
- [109] a) J. Rudolph, P. C. Sennhenn, C. P. Vlaar, K. B. Sharpless, *Angew. Chem. Int. Ed. Engl.* **1996**, *35*, 2810–2813; *Angew. Chem.* **1996**, *108*, 2991–2995; b) B. Tao, G. Schlingloff, K. B. Sharpless, *Tetrahedron Lett.* **1998**, *39*, 2507–2510; c) A. J. Morgan, C. E. Masse, J. S. Panek, *Org. Lett.* **1999**, *1*, 1949–1952.
- [110] K. Schmidt, W. Gunther, S. Stoyanova, B. Schubert, Z. Z. Li, M. Hamburger, *Org. Lett.* **2002**, *4*, 197–199.
- [111] K. Schmidt, U. Riese, Z. Z. Li, M. Hamburger, *J. Nat. Prod.* **2003**, *66*, 378–383.
- [112] Y. X. Cheng, B. Schneider, U. Riese, B. Schubert, Z. Z. Li, M. Hamburger, *J. Nat. Prod.* **2004**, *67*, 1854–1858.
- [113] a) M. Isaka, P. Chinthanom, S. Supothina, P. Tobwor, N. L. Hywel-Jones, *J. Nat. Prod.* **2010**, *73*, 2057–2060; b) H. J. Jessen, D. Barbaras, M. Hamburger, K. Gademann, *Org. Lett.* **2009**, *11*, 3446–3449; c) P. Burch, A. Chicca, J. Gertsch, K. Gademann, *ACS Med. Chem. Lett.* **2014**, *5*, 172–177.
- [114] a) H. J. Jessen, A. Schumacher, T. Shaw, A. Pfaltz, K. Gademann, *Angew. Chem. Int. Ed.* **2011**, *50*, 4222–4226; *Angew. Chem.* **2011**, *123*, 4308–4312; b) for a similar synthetic approach towards (+)-torrubiellone C, see H. J. Jessen, A. Schumacher, F. Schmid, A. Pfaltz, K. Gademann, *Org. Lett.* **2011**, *13*, 4368–4370.
- [115] F. Schmid, H. J. Jessen, P. Burch, K. Gademann, *Med. Chem. Commun.* **2013**, *4*, 135–139.
- [116] U. Riese, E. Ziegler, M. Hamburger, *FEBS Lett.* **2004**, *577*, 455–459.
- [117] P. Schröder, T. Förster, S. Kleine, C. Becker, A. Richters, S. Ziegler, D. Rauh, K. Kumar, H. Waldmann, *Angew. Chem. Int. Ed.* **2015**, *54*, 12398–12403.
- [118] a) A. G. Myers, S. B. Herzon, *J. Am. Chem. Soc.* **2003**, *125*, 12080–12081; b) S. B. Herzon, A. G. Myers, *J. Am. Chem. Soc.* **2005**, *127*, 5342–5344; c) J. E. Wulff, S. B. Herzon, R. Siegrist, A. G. Myers, *J. Am. Chem. Soc.* **2007**, *129*, 4898–4899; d) J. E. Wulff, R. Siegrist, A. G. Myers, *J. Am. Chem. Soc.* **2007**, *129*, 14444–14451.
- [119] a) B. M. Crowley, D. L. Boger, *J. Am. Chem. Soc.* **2006**, *128*, 2885–2892; b) for a more thorough review of this topic, see Ref. [4a].
- [120] a) U. N. Sundram, J. H. Griffin, T. I. Nicas, *J. Am. Chem. Soc.* **1996**, *118*, 13107–13108; b) A. Malabarba, T. I. Nicas, R. C. Thompson, *Med. Res. Rev.* **1997**, *17*, 69–137; c) J. P. Mackay, U. Gerhard, D. A. Beauregard, R. A. Maplestone, D. H. Williams, *J. Am. Chem. Soc.* **1994**, *116*, 4573–4580; d) J. P. Mackay, U. Gerhard, D. A. Beauregard, M. S. Westwell, M. S. Searle, D. H. Williams, *J. Am. Chem. Soc.* **1994**, *116*, 4581–4590.
- [121] a) W. Bauer, U. Briner, W. Doepfner, R. Haller, R. Huguenin, P. Marbach, T. J. Petcher, J. Pless, *Life Sci.* **1982**, *31*, 1133–1140; b) P. E. Battershill, S. P. Clissold, *Drugs* **1989**, *38*, 658–702.
- [122] a) L. Quadri, G. Bianchi, A. Cerri, G. Fedrizzi, P. Ferrari, M. Gobbi, P. Melloni, S. Sputore, M. Torri, *J. Med. Chem.* **1997**, *40*, 1561–1564; b) J. A. Staessen, T. Kuznetsova, R. Acceto, A. Bacchieri, E. Brand, M. Burnier, H. Celis, L. Citterio, P. W. de Leeuw, J. Filipovsky, A. Fournier, K. Kawecka-Jaszcz, P. Manunta, Y. Nikitin, E. T. O'Brien, J. Redon, L. Thijs, P. Ferrari, G. Valentini, G. Bianchi, *Pharmacogenomics* **2005**, *6*, 755–775; c) W. Schoner, G. Scheiner-Bobis, *Am. J. Physiol. Cell Physiol.* **2007**, *293*, C509–C536; d) P. Ferrari, M. Ferrandi, G. Valentini, P. Manunta, G. Bianchi, *Med. Hypotheses* **2007**, *68*, 1307–1314; e) P. Ferrari, *Biochim. Biophys. Acta Mol. Basis Dis.* **2010**, *1802*, 1254–1258; f) *First Patient Enrolled in Phase IIb Clinical Trial of Rostafuroxin in Italy*, press release, **2013**, available from: [http://www.leespharm.com/file/news/FOR%20IMMEDIATE%20RELEASE\\_Rostafuroxin%20first%20patient.pdf](http://www.leespharm.com/file/news/FOR%20IMMEDIATE%20RELEASE_Rostafuroxin%20first%20patient.pdf), last accessed: October 12, **2015**.
- [123] a) S. Zürcher, D. Wäckerlin, Y. Bethuel, B. Malisova, M. Textor, S. Tosatti, K. Gademann, *J. Am. Chem. Soc.* **2006**, *128*, 1064–1065; b) J. Y. Wach, S. Bonazzi, K. Gademann, *Angew. Chem. Int. Ed.* **2008**, *47*, 7123–7126; *Angew. Chem.* **2008**, *120*, 7232–7235; c) highlighted in: P. A. Wender, A. C. Donnelly, B. A. Loy, K. E. Near, D. Staveness in *Natural Products in Medicinal Chemistry*, 1st ed. (Ed.: S. Hanessian), Wiley-VCH, Weinheim, **2014**, pp. 475–543; and J. Y. Wach, K. Gademann, *Synlett* **2012**, 163–170.
- [124] a) K. C. Nicolaou, J. Tiebes, E. A. Theodorakis, F. P. J. T. Rutjes, K. Koide, M. Sato, E. Untersteller, *J. Am. Chem. Soc.* **1994**, *116*, 9371–9372; b) R. E. Gawley, K. S. Rein, G. Jeglitsch, D. J. Adams, E. A. Theodorakis, J. Tiebes, K. C. Nicolaou, D. G. Baden, *Chem. Biol.* **1995**, *2*, 533–541.
- [125] a) T. F. Molinski, D. J. Faulkner, C. H. He, G. D. Vanduyne, J. Clardy, *J. Org. Chem.* **1986**, *51*, 4564–4567; b) M. J. Schnermann, C. M. Beaudry, A. V. Egorova, R. S. Polishchuk, C. Sutterlin, L. E. Overman, *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 6158–6163.
- [126] G. H. Posner, P. M. O'Neill, *Acc. Chem. Res.* **2004**, *37*, 397–404.
- [127] E. J. Helfrich, S. Reiter, J. Piel, *Curr. Opin. Biotechnol.* **2014**, *29*, 107–115.
- [128] J. Piel, D. Hui, G. Wen, D. Butzke, M. Platzer, N. Fusetani, S. Matsunaga, *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 16222–16227.
- [129] a) M. S. Butler, A. A. B. Robertson, M. A. Cooper, *Nat. Prod. Rep.* **2014**, *31*, 1612–1661; b) for more thorough and recent reviews on natural product derived antibody–drug conjugates, see c) D. J. Newman, G. M. Cragg, *Mar. Drugs* **2014**, *12*, 255–278; d) R. V. J. Chari, M. L. Miller, W. C. Widdison, *Angew. Chem. Int. Ed.* **2014**, *53*, 3796–3827; *Angew. Chem.* **2014**, *126*, 3872–3904; e) H. P. Gerber, F. E. Koehn, R. T. Abraham, *Nat. Prod. Rep.* **2013**, *30*, 625–639; f) J. A. Flygare, T. H. Pillow, P. Aristoff, *Chem. Biol. Drug Des.* **2013**, *81*, 113–121; g) A. Mullard, *Nat. Rev. Drug Discovery* **2013**, *12*, 329–332; h) E. L. Sievers, P. D. Senter, *Annu. Rev. Med.* **2013**, *64*, 15–29.
- [130] G. T. Carter, *Nat. Prod. Rep.* **2011**, *28*, 1783–1789.
- [131] P. R. Hamann, L. M. Hinman, I. Hollander, C. F. Beyer, D. Lindh, R. Holcomb, W. Hallett, H. R. Tsou, J. Upešlaciš, D. Shochat, A. Mountain, D. A. Flowers, I. Bernstein, *Bioconjugate Chem.* **2002**, *13*, 47–58.
- [132] B. K. Shoichet, *Nat. Chem.* **2013**, *5*, 9–10.
- [133] B. Over, S. Wetzel, C. Grutter, Y. Nakai, S. Renner, D. Rauh, H. Waldmann, *Nat. Chem.* **2013**, *5*, 21–28.
- [134] a) S. Wetzel, R. S. Bon, K. Kumar, H. Waldmann, *Angew. Chem. Int. Ed.* **2011**, *50*, 10800–10826; *Angew. Chem.* **2011**,

- 123, 10990–11018; b) H. van Hattum, H. Waldmann, *J. Am. Chem. Soc.* **2014**, *136*, 11853–11859.
- [135] D. Reker, A. M. Perna, T. Rodrigues, P. Schneider, M. Reutlinger, B. Monch, A. Koeberle, C. Lamers, M. Gabler, H. Steinmetz, R. Muller, M. Schubert-Zsilavecz, O. Werz, G. Schneider, *Nat. Chem.* **2014**, *6*, 1072–1078.
- [136] T. Rodrigues, Y. C. Lin, M. Hartenfeller, S. Renner, Y. F. Lima, G. Schneider, *Chem. Commun.* **2015**, *51*, 7478–7481.
- [137] T. Miyao, D. Reker, P. Schneider, K. Funatsu, G. Schneider, *Planta Med.* **2015**, *81*, 429–435.
- [138] M. Pascolutti, M. Campitelli, B. Nguyen, N. Pham, A. D. Gorse, R. J. Quinn, *PLoS One* **2015**, *10*, e0120942.
- [139] J. Lanz, R. Riedl, *ChemMedChem* **2015**, *10*, 451–454.

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